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(57) Abstract

PDGF-D, a new member of the PDGF/VEGF family of growth factors, is described, as well as the nucleotide sequence encoding it, methods for producing it, antibodies and other antagonists to it, transfected and transformed host cells expressing it, pharmaceutical compositions containing it, and uses thereof in medical and diagnostic applications.

PLATELET-DERIVED GROWTH FACTOR D,
DNA CODING THEREFOR, AND USES THEREOF

This invention relates to growth factors for cells expressing receptors to a novel growth factor that include endothelial cells, connective tissue cells (such as fibroblasts) myofibroblasts and glial cells, and in particular to a novel platelet-derived growth factor/vascular endothelial growth factor-like growth factor, a polynucleotide sequence encoding the factor, and to pharmaceutical and diagnostic compositions and methods utilizing or derived from the factor.

10 BACKGROUND OF THE INVENTION

In the developing embryo, the primary vascular network is established by *in situ* differentiation of mesodermal cells in a process called vasculogenesis. It is believed that all subsequent processes involving the generation of new vessels in the embryo and neovascularization in adults, are governed by the sprouting or splitting of new capillaries from the pre-existing vasculature in a process called angiogenesis (Pepper et al., Enzyme & Protein, 1996 49 138-162; Breier et al., Dev. Dyn. 1995 204 228-239; Risau, Nature, 1997 386 671-674). Angiogenesis is not only involved in embryonic development and normal tissue growth, repair, and regeneration, but is also involved in the female reproductive cycle, establishment and maintenance of pregnancy, and in repair of wounds and fractures. In addition to angiogenesis which takes place in the normal individual, angiogenic events are involved in a number of pathological processes, notably tumor growth and metastasis, and other conditions in which blood vessel proliferation, especially of the microvascular system, is increased, such as diabetic retinopathy, psoriasis and arthropathies. Inhibition of angiogenesis is useful in preventing or alleviating these pathological processes.

On the other hand, promotion of angiogenesis is desirable in situations where vascularization is to be established or extended, for example after tissue or organ transplantation, or to stimulate establishment of collateral circulation in tissue infarction or arterial stenosis, such as in coronary heart disease and thromboangitis obliterans.

The angiogenic process is highly complex and involves the maintenance of the endothelial cells in the cell cycle, degradation of the extracellular matrix, migration and invasion of the surrounding tissue and finally, tube formation. The molecular mechanisms underlying the complex angiogenic processes are far from being understood.

Because of the crucial role of angiogenesis in so many physiological and pathological processes, factors involved in the control of angiogenesis have been intensively investigated. A number of growth factors have been shown to be involved in the regulation of angiogenesis; these include fibroblast growth factors (FGFs), platelet-derived growth factor (PDGF), transforming growth factor alpha (TGF α), and hepatocyte growth factor (HGF). See for example Folkman et al., J. Biol. Chem., 1992 267 10931-10934 for a review.

It has been suggested that a particular family of endothelial cell-specific growth factors, the vascular endothelial growth factors (VEGFs), and their corresponding receptors is primarily responsible for stimulation of endothelial cell growth and differentiation, and for certain functions of the differentiated cells. These factors are members of the PDGF family, and appear to act primarily via endothelial receptor tyrosine kinases (RTKs).

Nine different proteins have been identified in the PDGF family, namely two PDGFs (A and B), VEGF and six members that are closely related to VEGF. The six members closely related to VEGF are: VEGF-B, described in International Patent Application PCT/US96/02957 (WO 96/26736) and in U.S. Patents 5,840,693 and

5,607,918 by Ludwig Institute for Cancer Research and The University of Helsinki; VEGF-C, described in Joukov et al., EMBO J., 1996 15 290-298 and Lee et al., Proc. Natl. Acad. Sci. USA, 1996 93 1988-1992; VEGF-D, described in International Patent Application No. PCT/US97/14696 (WO 98/07832), and Achen et al., Proc. Natl. Acad. Sci. USA, 1998 95 548-553; the placenta growth factor (PlGF), described in Maglione et al., Proc. Natl. Acad. Sci. USA, 1991 88 9267-9271; VEGF2, described in International Patent Application No. PCT/US94/05291 (WO 95/24473) by Human Genome Sciences, Inc; and VEGF3, described in International Patent Application No. PCT/US95/07283 (WO 96/39421) by Human Genome Sciences, Inc. Each VEGF family member has between 30% and 45% amino acid sequence identity with VEGF. The VEGF family members share a VEGF homology domain which contains the six cysteine residues which form the cysteine knot motif. Functional characteristics of the VEGF family include varying degrees of mitogenicity for endothelial cells, induction of vascular permeability and angiogenic and lymphangiogenic properties.

Vascular endothelial growth factor (VEGF) is a homodimeric glycoprotein that has been isolated from several sources. VEGF shows highly specific mitogenic activity for endothelial cells. VEGF has important regulatory functions in the formation of new blood vessels during embryonic vasculogenesis and in angiogenesis during adult life (Carmeliet et al., Nature, 1996 380 435-439; Ferrara et al., Nature, 1996 380 439-442; reviewed in Ferrara and Davis-Smyth, Endocrine Rev., 1997 18 4-25). The significance of the role played by VEGF has been demonstrated in studies showing that inactivation of a single VEGF allele results in embryonic lethality due to failed development of the vasculature (Carmeliet et al., Nature, 1996 380 435-439; Ferrara et al., Nature, 1996 380 439-442). In addition VEGF has strong chemoattractant activity towards monocytes, can induce the plasminogen activator and the plasminogen activator inhibitor in endothelial cells, and can also

induce microvascular permeability. Because of the latter activity, it is sometimes referred to as vascular permeability factor (VPF). The isolation and properties of VEGF have been reviewed; see Ferrara et al., J. Cellular Biochem., 1991 47 211-218 and Connolly, J. Cellular Biochem., 1991 47 219-223. Alternative mRNA splicing of a single VEGF gene gives rise to five isoforms of VEGF.

VEGF-B has similar angiogenic and other properties to those of VEGF, but is distributed and expressed in tissues differently from VEGF. In particular, VEGF-B is very strongly expressed in heart, and only weakly in lung, whereas the reverse is the case for VEGF. This suggests that VEGF and VEGF-B, despite the fact that they are co-expressed in many tissues, may have functional differences.

VEGF-B was isolated using a yeast co-hybrid interaction trap screening technique by screening for cellular proteins which might interact with cellular retinoid acid-binding protein type I (CRABP-I). Its isolation and characteristics are described in detail in PCT/US96/02957 and in Olofsson et al., Proc. Natl. Acad. Sci. USA, 1996 93 2576-2581.

VEGF-C was isolated from conditioned media of the PC-3 prostate adenocarcinoma cell line (CRL1435) by screening for ability of the medium to produce tyrosine phosphorylation of the endothelial cell-specific receptor tyrosine kinase VEGFR-3 (Flt4), using cells transfected to express VEGFR-3. VEGF-C was purified using affinity chromatography with recombinant VEGFR-3, and was cloned from a PC-3 cDNA library. Its isolation and characteristics are described in detail in Joukov et al., EMBO J., 1996 15 290-298.

VEGF-D was isolated from a human breast cDNA library, commercially available from Clontech, by screening with an expressed sequence tag obtained from a human cDNA library designated "Soares Breast 3NbHBst" as a hybridization probe (Achen

et al., Proc. Natl. Acad. Sci. USA, 1998 95 548-553). Its isolation and characteristics are described in detail in International Patent Application No. PCT/US97/14696 (WO98/07832).

5 The VEGF-D gene is broadly expressed in the adult human, but is certainly not ubiquitously expressed. VEGF-D is strongly expressed in heart, lung and skeletal muscle. Intermediate levels of VEGF-D are expressed in spleen, ovary, small intestine and colon, and a lower expression occurs in kidney, pancreas, thymus, prostate and testis. No VEGF-D mRNA was detected in RNA from
10 brain, placenta, liver or peripheral blood leukocytes.

PlGF was isolated from a term placenta cDNA library. Its isolation and characteristics are described in detail in Maglione et al., Proc. Natl. Acad. Sci. USA, 1991 88 9267-9271. Presently its biological function is not well understood.

15 VEGF2 was isolated from a highly tumorigenic, oestrogen-independent human breast cancer cell line. While this molecule is stated to have about 22% homology to PDGF and 30% homology to VEGF, the method of isolation of the gene encoding VEGF2 is unclear, and no characterization of the biological activity is
20 disclosed.

VEGF3 was isolated from a cDNA library derived from colon tissue. VEGF3 is stated to have about 36% identity and 66% similarity to VEGF. The method of isolation of the gene encoding VEGF3 is unclear and no characterization of the biological
25 activity is disclosed.

Similarity between two proteins is determined by comparing the amino acid sequence and conserved amino acid substitutions of one of the proteins to the sequence of the second protein, whereas identity is determined without including the conserved amino acid
30 substitutions.

PDGF/VEGF family members act primarily by binding to receptor tyrosine kinases. Five endothelial cell-specific receptor tyrosine kinases have been identified, namely VEGFR-1 (Flt-1),

VEGFR-2 (KDR/Flk-1), VEGFR-3 (Flt4), Tie and Tek/Tie-2. All of these have the intrinsic tyrosine kinase activity which is necessary for signal transduction. The essential, specific role in vasculogenesis and angiogenesis of VEGFR-1, VEGFR-2, VEGFR-3, Tie and Tek/Tie-2 has been demonstrated by targeted mutations inactivating these receptors in mouse embryos.

The only receptor tyrosine kinases known to bind VEGFs are VEGFR-1, VEGFR-2 and VEGFR-3. VEGFR-1 and VEGFR-2 bind VEGF with high affinity, and VEGFR-1 also binds VEGF-B and PlGF. VEGF-C has been shown to be the ligand for VEGFR-3, and it also activates VEGFR-2 (Joukov et al., The EMBO Journal, 1996 15 290-298). VEGF-D binds to both VEGFR-2 and VEGFR-3. A ligand for Tek/Tie-2 has been described in International Patent Application No. PCT/US95/12935 (WO 96/11269) by Regeneron Pharmaceuticals, Inc. The ligand for Tie has not yet been identified.

Recently, a novel 130-135 kDa VEGF isoform specific receptor has been purified and cloned (Soker et al., Cell, 1998 92 735-745). The VEGF receptor was found to specifically bind the VEGF₁₆₅ isoform via the exon 7 encoded sequence, which shows weak affinity for heparin (Soker et al., Cell, 1998 92 735-745). Surprisingly, the receptor was shown to be identical to human neuropilin-1 (NP-1), a receptor involved in early stage neuromorphogenesis. PlGF-2 also appears to interact with NP-1 (Migdal et al., J. Biol. Chem., 1998 273 22272-22278).

VEGFR-1, VEGFR-2 and VEGFR-3 are expressed differently by endothelial cells. Both VEGFR-1 and VEGFR-2 are expressed in blood vessel endothelia (Oelrichs et al., Oncogene, 1992 8 11-18; Kaipainen et al., J. Exp. Med., 1993 178 2077-2088; Dumont et al., Dev. Dyn., 1995 203 80-92; Fong et al., Dev. Dyn., 1996 207 1-10) and VEGFR-3 is mostly expressed in the lymphatic endothelium of adult tissues (Kaipainen et al., Proc. Natl. Acad. Sci. USA, 1995 9 3566-3570). VEGFR-3 is also expressed in the blood vasculature surrounding tumors.

Disruption of the VEGFR genes results in aberrant development of the vasculature leading to embryonic lethality around midgestation. Analysis of embryos carrying a completely inactivated VEGFR-1 gene suggests that this receptor is required for functional organization of the endothelium (Fong et al., Nature, 1995 376 66-70). However, deletion of the intracellular tyrosine kinase domain of VEGFR-1 generates viable mice with a normal vasculature (Hiratsuka et al., Proc. Natl. Acad. Sci. USA 1998 95 9349-9354). The reasons underlying these differences remain to be explained but suggest that receptor signalling via the tyrosine kinase is not required for the proper function of VEGFR-1. Analysis of homozygous mice with inactivated alleles of VEGFR-2 suggests that this receptor is required for endothelial cell proliferation, hematopoiesis and vasculogenesis (Shalaby et al., Nature, 1995 376 62-66; Shalaby et al., Cell, 1997 89 981-990). Inactivation of VEGFR-3 results in cardiovascular failure due to abnormal organization of the large vessels (Dumont et al. Science, 1998 282 946-949).

Although VEGFR-1 is mainly expressed in endothelial cells during development, it can also be found in hematopoietic precursor cells during early stages of embryogenesis (Fong et al., Nature, 1995 376 66-70). It is also expressed by most, if not all, vessels in embryos (Breier et al., Dev. Dyn., 1995 204 228-239; Fong et al., Dev. Dyn., 1996 207 1-10). In adults, monocytes and macrophages also express this receptor (Barleon et al., Blood, 1996 87 3336-3343).

The receptor VEGFR-3 is widely expressed on endothelial cells during early embryonic development, but as embryogenesis proceeds, it becomes restricted to venous endothelium and then to the lymphatic endothelium (Kaipainen et al., Cancer Res., 1994 54 6571-6577; Kaipainen et al., Proc. Natl. Acad. Sci. USA, 1995 92 3566-3570). VEGFR-3 continues to be expressed on lymphatic endothelial cells in adults. This receptor is essential for

vascular development during embryogenesis. Targeted inactivation of both copies of the VEGFR-3 gene in mice resulted in defective blood vessel formation characterized by abnormally organized large vessels with defective lumens, leading to fluid accumulation in the pericardial cavity and cardiovascular failure at post-coital day 9.5. On the basis of these findings it has been proposed that VEGFR-3 is required for the maturation of primary vascular networks into larger blood vessels. However, the role of VEGFR-3 in the development of the lymphatic vasculature could not be studied in these mice because the embryos died before the lymphatic system emerged. Nevertheless it is assumed that VEGFR-3 plays a role in development of the lymphatic vasculature and lymphangiogenesis given its specific expression in lymphatic endothelial cells during embryogenesis and adult life. This is supported by the finding that ectopic expression of VEGF-C, a ligand for VEGFR-3, in the skin of transgenic mice, resulted in lymphatic endothelial cell proliferation and vessel enlargement in the dermis. Furthermore this suggests that VEGF-C may have a primary function in lymphatic endothelium, and a secondary function in angiogenesis and permeability regulation which is shared with VEGF (Joukov et al., EMBO J., 1996 15 290-298).

Some inhibitors of the VEGF/VEGF-receptor system have been shown to prevent tumor growth via an anti-angiogenic mechanism; see Kim et al., Nature, 1993 362 841-844 and Saleh et al., Cancer Res., 1996 56 393-401.

As mentioned above, the VEGF family of growth factors are members of the PDGF family. PDGF plays a important role in the growth and/or motility of connective tissue cells, fibroblasts, myofibroblasts and glial cells (Heldin et al., "Structure of platelet-derived growth factor: Implications for functional properties", Growth Factor, 1993 8 245-252). In adults, PDGF stimulates wound healing (Robson et al., Lancet, 1992 339 23-25). Structurally, PDGF isoforms are disulfide-bonded dimers of

homologous A- and B-polypeptide chains, arranged as homodimers (PDGF-AA and PDGF-BB) or a heterodimer (PDGF-AB).

PDGF isoforms exert their effects on target cells by binding to two structurally related receptor tyrosine kinases (RTKs). The alpha-receptor binds both the A- and B-chains of PDGF, whereas the beta-receptor binds only the B-chain. These two receptors are expressed by many *in vitro* grown cell lines, and are mainly expressed by mesenchymal cells *in vivo*. The PDGFs regulate cell proliferation, cell survival and chemotaxis of many cell types *in vitro* (reviewed in Heldin et al., *Biochim Biophys Acta.*, 1998 1378 F79-113). *In vivo*, they exert their effects in a paracrine mode since they often are expressed in epithelial (PDGF-A) or endothelial cells (PDGF-B) in close apposition to the PDGFR expressing mesenchyme. In tumor cells and in cell lines grown *in vitro*, coexpression of the PDGFs and the receptors generate autocrine loops which are important for cellular transformation (Betsholtz et al., *Cell*, 1984 39 447-57; Keating et al., *J. R. Coll Surg Edinb.*, 1990 35 172-4). Overexpression of the PDGFs have been observed in several pathological conditions, including malignancies, arteriosclerosis, and fibroproliferative diseases (reviewed in Heldin et al., *The Molecular and Cellular Biology of Wound Repair*, New York: Plenum Press, 1996, 249-273).

The importance of the PDGFs as regulators of cell proliferation and survival are well illustrated by recent gene targeting studies in mice that have shown distinct physiological roles for the PDGFs and their receptors despite the overlapping ligand specificities of the PDGFRs. Homozygous null mutations for either of the two PDGF ligands or the receptors are lethal. Approximately 50% of the homozygous PDGF-A deficient mice have an early lethal phenotype, while the surviving animals have a complex postnatal phenotype with lung emphysema due to improper alveolar septum formation because of a lack of alveolar myofibroblasts (Boström et al., *Cell*, 1996 85 863-873). The PDGF-A deficient

mice also have a dermal phenotype characterized by thin dermis, misshapen hair follicles and thin hair (Karlsson et al., Development, 1999 126 2611-2). PDGF-A is also required for normal development of oligodendrocytes and subsequent myelination of the central nervous system (Fruttiger et al., Development, 1999 126 457-67). The phenotype of PDGFR-alpha deficient mice is more severe with early embryonic death at E10, incomplete cephalic closure, impaired neural crest development, cardiovascular defects, skeletal defects, and odemas [Soriano et al., Development, 1997 124 2691-70). The PDGF-B and PDGFR-beta deficient mice develop similar phenotypes that are characterized by renal, hematological and cardiovascular abnormalities (Levéen et al., Genes Dev., 1994 8 1875-1887; Soriano et al., Genes Dev., 1994 8 1888-96; Lindahl et al., Science, 1997 277 242-5; Lindahl, Development, 1998 125 3313-2), where the renal and cardiovascular defects, at least in part, are due to the lack of proper recruitment of mural cells (vascular smooth muscle cells, pericytes or mesangial cells) to blood vessels (Levéen et al., Genes Dev., 1994 8 1875-1887; Lindahl et al., Science, 1997 277 242-5; Lindahl et al., Development, 1998 125 3313-2).

SUMMARY OF THE INVENTION

The invention generally provides an isolated novel growth factor which has the ability to stimulate and/or enhance proliferation or differentiation and/or growth and/or motility of cells expressing a PDGF-D receptor including, but not limited to, endothelial cells, connective tissue cells, myofibroblasts and glial cells, an isolated polynucleotide sequence encoding the novel growth factor, and compositions useful for diagnostic and/or therapeutic applications.

According to one aspect, the invention provides an isolated and purified nucleic acid molecule which comprises a

polynucleotide sequence having at least 85% identity, more preferably at least 90%, and most preferably at least 95% identity to at least nucleotides 1 to 600 of the sequence set out in Figure 3 (SEQ ID NO:3), at least nucleotides 1 to 966 of the sequence set out in Figure 5 (SEQ ID NO:5), at least nucleotides 176 to 1288 of the sequence set out in Figure 7 (SEQ ID NO:7) or at least nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7). The sequence of at least nucleotides 1 to 600 of the sequence set out in Figure 3 or at least nucleotides 1 to 966 of the sequence set out in Figure 5 encodes a 5'-truncated polypeptide, designated PDGF-D (formally designated "VEGF-G"), while at least nucleotides 173 to 1288 of the sequence set out in Figure 7 (SEQ ID NO:7) encodes a full-length PDGF-D. PDGF-D is structurally homologous to PDGF-A, PDGF-B, VEGF, VEGF-B, VEGF-C and VEGF-D. The sequence of at least nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7) encodes a portion of the PDGF/VEGF homology domain, which is the bioactive fragment of PDGF-D. This bioactive fragment would also be encoded by the sequence of at least nucleotides 1 to 600 of the sequence set out in Figure 3 or at least nucleotides 1 to 966 of the sequence set out in Figure 5. In a preferred embodiment, the nucleic acid molecule is a cDNA which comprises at least nucleotides 1 to 600 of the sequence set out in Figure 3 (SEQ ID NO:3), at least nucleotides 1 to 966 of the sequence set out in Figure 5 (SEQ ID NO:5), at least nucleotides 173 to 1288 of the sequence set out in Figure 7 (SEQ ID NO:7) or at least nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7). This aspect of the invention also encompasses DNA molecules having a sequence such that they hybridize under stringent conditions with at least nucleotides 1 to 600 of the sequence set out in Figure 3 (SEQ ID NO:3), at least nucleotides 1 to 966 of the sequence set out in Figure 5 (SEQ ID NO:5), at least nucleotides 173 to 1288 of the sequence set out in Figure 7 (SEQ ID NO:7) or at least

nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7) or fragments thereof.

According to a second aspect, the polypeptide of the invention has the ability to stimulate and/or enhance proliferation and/or differentiation and/or growth and/or motility of cells expressing a PDGF-C receptor including, but not limited to, endothelial cells, connective tissue cells, myofibroblasts and glial cells and comprises a sequence of amino acids corresponding to the amino acid sequence set out in Figure 4 (SEQ ID NO:4) or Figure 6 (SEQ ID NO:6), or Figure 8 (SEQ ID NO:8), or a fragment or analog thereof which has the ability to stimulate endothelial cell proliferation, differentiation, migration and/or survival of and/or growth and/or motility of connective tissue cells (such as fibroblasts), myofibroblasts and glial cells. Preferably the polypeptides have at least 85% identity, more preferably at least 90%, and most preferably at least 95% identity to the amino acid sequence of in Figure 4 (SEQ ID NO:4) or Figure 6 (SEQ ID NO:6) or Figure 8 (SEQ ID NO:8), or a fragment or analog thereof having the biological activity of PDGF-D. A preferred fragment is a truncated form of PDGF-D comprising a portion of the PDGF/VEGF homology domain (PVHD) of PDGF-D. The portion of the PVHD is from residues 255-371 of Figure 8 where the putative proteolytic processing site RKSK starts at amino acid residue 255 (SEQ ID NO:8). However, the PVHD extends toward the N terminus up to residue 235 of Figure 8 (SEQ ID NO:8). Herein the PVHD is defined as truncated PDGF-D. The truncated PDGF-D is the putative activated form of PDGF-D.

As used in this application, percent sequence identity is determined by using the alignment tool of "MEGALIGN" from the Lasergene package (DNASTAR, Ltd. Abacus House, Manor Road, West Ealing, London W130AS United Kingdom). The MEGALIGN is based on the J. Hein method (Methods in Enzymology, 1990 183 626-645). The PAM 250 residue weight table is used with a gap penalty of eleven

and a gap length penalty of three and a K-tuple value of two in the pairwise alignments. The alignment is then refined manually, and the number of identities are estimated in the regions available for a comparison.

5 Preferably the polypeptide or the encoded polypeptide from a polynucleotide has the ability to stimulate one or more of proliferation, differentiation, motility, survival or vascular permeability of cells expressing a PDGF-D receptor including, but not limited to, vascular endothelial cells, lymphatic endothelial
10 cells, connective tissue cells (such as fibroblasts), myofibroblasts and glial cells. Preferably the polypeptide or the encoded polypeptide from a polynucleotide has the ability to stimulate wound healing. PDGF-D can also have antagonistic effects on cells, but are included in the biological activities
15 of PDGF-D. These abilities are referred to hereinafter as "biological activities of PDGF-D" and can be readily tested by methods known in the art.

As used herein, the term "PDGF-D" collectively refers to the polypeptides of Figure 4 (SEQ ID NO:4), Figure 6 (SEQ ID NO:6) or
20 Figure 8 (SEQ ID NO:8), and fragments or analogs thereof which have the biological activity of PDGF-D as defined above, and to a polynucleotide which can code for PDGF-D, or a fragment or analog thereof having the biological activity of PDGF-D. The polynucleotide can be naked and/or in a vector or liposome.

25 In another preferred aspect, the invention provides a polypeptide possessing an amino acid sequence:

PXCLLVXRCGGNCXC (SEQ ID NO:25)

which is unique to PDGF-D and differs from the other members of the PDGF/VEGF family of growth factors because of the insertion
30 of the three amino acid residues (NCG) between the third and fourth cysteines (see Figure 9 - SEQ ID NOS:10-18).

Polypeptides comprising conservative substitutions, insertions, or deletions, but which still retain the biological

activity of PDGF-D are clearly to be understood to be within the scope of the invention. Persons skilled in the art will be well aware of methods which can readily be used to generate such polypeptides, for example the use of site-directed mutagenesis, or specific enzymatic cleavage and ligation. The skilled person will also be aware that peptidomimetic compounds or compounds in which one or more amino acid residues are replaced by a non-naturally occurring amino acid or an amino acid analog may retain the required aspects of the biological activity of PDGF-D. Such compounds can readily be made and tested for their ability to show the biological activity of PDGF-D by routine activity assay procedures such as the fibroblast proliferation assay and are also within the scope of the invention.

In addition, possible variant forms of the PDGF-D polypeptide which may result from alternative splicing, as are known to occur with VEGF and VEGF-B, and naturally-occurring allelic variants of the nucleic acid sequence encoding PDGF-D are encompassed within the scope of the invention. Allelic variants are well known in the art, and represent alternative forms or a nucleic acid sequence which comprise substitution, deletion or addition of one or more nucleotides, but which do not result in any substantial functional alteration of the encoded polypeptide.

Such variant forms of PDGF-D can be prepared by targeting non-essential regions of the PDGF-D polypeptide for modification. These non-essential regions are expected to fall outside the strongly-conserved regions indicated in Figure 9 (SEQ ID NOs:10-18). In particular, the growth factors of the PDGF family, including PDGF-D, are dimeric. PDGF-D differs slightly from VEGF, VEGF-B, VEGF-C, VEGF-D, PlGF, PDGF-A and PDGF-B because it shows complete conservation of only seven of the eight cysteine residues in the PVHD (Olofsson et al., Proc. Natl. Acad. Sci. USA, 1996 93 2576-2581; Joukov et al., EMBO J., 1996 15 290-298). These cysteines are thought to be involved in intra- and inter-molecular

disulfide bonding. Loops 1, 2 and 3 of each subunit, which are formed by intra-molecular disulfide bonding, are involved in binding to the receptors for the PDGF/VEGF family of growth factors (Andersson et al., Growth Factors, 1995 12 159-164).

5 Persons skilled in the art thus are well aware that these cysteine residues should be preserved in any proposed variant form, and that the active sites present in loops 1, 2 and 3 also should be preserved. However, other regions of the molecule can be expected to be of lesser importance for biological function, and therefore offer suitable targets for modification. Modified polypeptides can readily be tested for their ability to show the biological activity of PDGF-D by routine activity assay procedures such as the fibroblast proliferation assay.

10 It is contemplated that some modified PDGF-D polypeptides will have the ability to bind to PDGF-D receptors on cells including, but not limited to, endothelial cells, connective tissue cells, myofibroblasts and/or glial cells, but will be unable to stimulate cell proliferation, differentiation, migration, motility or survival or to induce vascular proliferation, connective tissue development or wound healing. These modified polypeptides are expected to be able to act as competitive or non-competitive inhibitors of the PDGF-D polypeptides and growth factors of the PDGF/VEGF family, and to be useful in situations where prevention or reduction of the PDGF-D polypeptide or PDGF/VEGF family growth factor action is desirable. Thus such receptor-binding but non-mitogenic, non-differentiation inducing, non-migration inducing, non-motility inducing, non-survival promoting, non-connective tissue promoting, non-wound healing or non-vascular proliferation inducing variants of the PDGF-D polypeptide are also within the scope of the invention, and are referred to herein as "receptor-binding but otherwise inactive variants". Because PDGF-D forms a dimer in order to activate its only known receptor, it is contemplated that

one monomer comprises the receptor-binding but otherwise inactive variant modified PDGF-D polypeptide and a second monomer comprises a wild-type PDGF-D or a wild-type growth factor of the PDGF/VEGF family. These dimers can bind to its corresponding receptor but cannot induce downstream signaling.

It is also contemplated that there are other modified PDGF-D polypeptides that can prevent binding of a wild-type PDGF-D or a wild-type growth factor of the PDGF/VEGF family to its corresponding receptor on cells including, but not limited to, endothelial cells, connective tissue cells (such as fibroblasts), myofibroblasts and/or glial cells. Thus these dimers will be unable to stimulate endothelial cell proliferation, differentiation, migration, survival, or induce vascular permeability, and/or stimulate proliferation and/or differentiation and/or motility of connective tissue cells, myofibroblasts or glial cells. These modified polypeptides are expected to be able to act as competitive or non-competitive inhibitors of the PDGF-D growth factor or a growth factor of the PDGF/VEGF family, and to be useful in situations where prevention or reduction of the PDGF-D growth factor or PDGF/VEGF family growth factor action is desirable. Such situations include the tissue remodeling that takes place during invasion of tumor cells into a normal cell population by primary or metastatic tumor formation. Thus such the PDGF-D or PDGF/VEGF family growth factor-binding but non-mitogenic, non-differentiation inducing, non-migration inducing, non-motility inducing, non-survival promoting, non-connective tissue promoting, non-wound healing or non-vascular proliferation inducing variants of the PDGF-D growth factor are also within the scope of the invention, and are referred to herein as "the PDGF-D growth factor-dimer forming but otherwise inactive or interfering variants".

An example of a PDGF-D growth factor-dimer forming but otherwise inactive or interfering variant is where the PDGF-D has

a mutation which prevents cleavage of CUB domain from the protein. It is further contemplated that a PDGF-D growth factor-dimer forming but otherwise inactive or interfering variant could be made to comprise a monomer, preferably an activated monomer, of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-C, PDGF-A, PDGF-B, PDGF-C, PDGF-D or PlGF linked to a CUB domain that has a mutation which prevents cleavage of CUB domain from the protein. Dimers formed with the above mentioned PDGF-D growth factor-dimer forming but otherwise inactive or interfering variants and the monomers linked to the mutant CUB domain would be unable to bind to their corresponding receptors.

A variation on this contemplation would be to insert a proteolytic site between an activated monomer of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-C, PDGF-A, PDGF-B, PDGF-C, PDGF-D or PlGF and the mutant CUB domain linkage which is dimerized to an activated monomer of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-A, PDGF-B, PDGF-C, PDGF-D or PlGF. An addition of the specific protease(s) for this proteolytic site would cleave the CUB domain and thereby release an activated dimer that can then bind to its corresponding receptor. In this way, a controlled release of an activated dimer is made possible.

According to a third aspect, the invention provides a purified and isolated nucleic acid encoding a polypeptide or polypeptide fragment of the invention as defined above. The nucleic acid may be DNA, genomic DNA, cDNA or RNA, and may be single-stranded or double stranded. The nucleic acid may be isolated from a cell or tissue source, or of recombinant or synthetic origin. Because of the degeneracy of the genetic code, the person skilled in the art will appreciate that many such coding sequences are possible, where each sequence encodes the amino acid sequence shown in Figure 4 (SEQ ID NO:4), Figure 6 (SEQ ID NO:6) or Figure 8 (SEQ ID NO:8), a bioactive fragment or analog thereof, a receptor-binding but otherwise inactive or partially

inactive variant thereof or a PDGF-D dimer-forming but otherwise inactive or interfering variants thereof.

5 A fourth aspect of the invention provides vectors comprising the cDNA of the invention or a nucleic acid molecule according to the third aspect of the invention, and host cells transformed or
transfected with nucleic acids molecules or vectors of the invention. These may be eukaryotic or prokaryotic in origin. These cells are particularly suitable for expression of the polypeptide of the invention, and include insect cells such as Sf9
10 cells, obtainable from the American Type Culture Collection (ATCC SRL-171), transformed with a baculovirus vector, and the human embryo kidney cell line 293-EBNA transfected by a suitable expression plasmid. Preferred vectors of the invention are expression vectors in which a nucleic acid according to the
15 invention is operatively connected to one or more appropriate promoters and/or other control sequences, such that appropriate host cells transformed or transfected with the vectors are capable of expressing the polypeptide of the invention. Other preferred
vectors are those suitable for transfection of mammalian cells, or for gene therapy, such as adenoviral-, vaccinia- or retroviral-
20 based vectors or liposomes. A variety of such vectors is known in the art.

The invention also provides a method of making a vector capable of expressing a polypeptide encoded by a nucleic acid
25 molecule according to the invention, comprising the steps of operatively connecting the nucleic acid molecule to one or more appropriate promoters and/or other control sequences, as described above.

The invention further provides a method of making a
30 polypeptide according to the invention, comprising the steps of expressing a nucleic acid or vector of the invention in a host cell, and isolating the polypeptide from the host cell or from the host cell's growth medium.

In yet a further aspect, the invention provides an antibody specifically reactive with a polypeptide of the invention or a fragment of the polypeptide. This aspect of the invention includes antibodies specific for the variant forms, immunoreactive fragments, analogs and recombinants of PDGF-D. Such antibodies are useful as inhibitors or agonists of PDGF-D and as diagnostic agents for detecting and quantifying PDGF-D. Polyclonal or monoclonal antibodies may be used. Monoclonal and polyclonal antibodies can be raised against polypeptides of the invention or fragment or analog thereof using standard methods in the art. In addition the polypeptide can be linked to an epitope tag, such as the FLAG[®] octapeptide (Sigma, St. Louis, MO), to assist in affinity purification. For some purposes, for example where a monoclonal antibody is to be used to inhibit effects of PDGF-D in a clinical situation, it may be desirable to use humanized or chimeric monoclonal antibodies. Such antibodies may be further modified by addition of cytotoxic or cytostatic drug(s). Methods for producing these, including recombinant DNA methods, are also well known in the art.

This aspect of the invention also includes an antibody which recognizes PDGF-D and is suitably labeled.

Polypeptides or antibodies according to the invention may be labeled with a detectable label, and utilized for diagnostic purposes. Similarly, the thus-labeled polypeptide of the invention may be used to identify its corresponding receptor *in situ*. The polypeptide or antibody may be covalently or non-covalently coupled to a suitable supermagnetic, paramagnetic, electron dense, ecogenic or radioactive agent for imaging. For use in diagnostic assays, radioactive or non-radioactive labels may be used. Examples of radioactive labels include a radioactive atom or group, such as ¹²⁵I or ³²P. Examples of non-radioactive labels include enzymatic labels, such as horseradish peroxidase or fluorimetric labels, such as fluorescein-5-isothiocyanate

(FITC). Labeling may be direct or indirect, covalent or non-covalent.

Clinical applications of the invention include diagnostic applications, acceleration of angiogenesis in tissue or organ transplantation, or stimulation of wound healing, or connective tissue development, or to establish collateral circulation in tissue infarction or arterial stenosis, such as coronary artery disease, and inhibition of angiogenesis in the treatment of cancer or of diabetic retinopathy and inhibition of tissue remodeling that takes place during invasion of tumor cells into a normal cell population by primary or metastatic tumor formation. Quantitation of PDGF-D in cancer biopsy specimens may be useful as an indicator of future metastatic risk.

PDGF-D may also be relevant to a variety of lung conditions. PDGF-D assays could be used in the diagnosis of various lung disorders. PDGF-D could also be used in the treatment of lung disorders to improve blood circulation in the lung and/or gaseous exchange between the lungs and the blood stream. Similarly, PDGF-D could be used to improve blood circulation to the heart and O₂ gas permeability in cases of cardiac insufficiency. In a like manner, PDGF-D could be used to improve blood flow and gaseous exchange in chronic obstructive airway diseases.

Thus the invention provides a method of stimulation of angiogenesis, lymphangiogenesis, neovascularization, connective tissue development and/or wound healing in a mammal in need of such treatment, comprising the step of administering an effective dose of PDGF-D, or a fragment or an analog thereof which has the biological activity of PDGF-D to the mammal. Optionally the PDGF-D, or fragment or analog thereof may be administered together with, or in conjunction with, one or more of VEGF, VEGF-B, VEGF-C, VEGF-D, PlGF, PDGF-A, PDGF-B, PDGF-C, FGF and/or heparin.

Conversely, PDGF-D antagonists (e.g. antibodies and/or competitive or noncompetitive inhibitors of binding of PDGF-D in

both dimer formation and receptor binding) could be used to treat conditions, such as congestive heart failure, involving accumulation of fluid in, for example, the lung resulting from increases in vascular permeability, by exerting an offsetting effect on vascular permeability in order to counteract the fluid accumulation. Administrations of PDGF-D could be used to treat malabsorptive syndromes in the intestinal tract, liver or kidneys as a result of its blood circulation increasing and vascular permeability increasing activities.

Thus, the invention provides a method of inhibiting angiogenesis, lymphangiogenesis, neovascularization, connective tissue development and/or wound healing in a mammal in need of such treatment, comprising the step of administering an effective amount of an antagonist of PDGF-D to the mammal. The antagonist may be any agent that prevents the action of PDGF-D, either by preventing the binding of PDGF-D to its corresponding receptor on the target cell, or by preventing activation of the receptor, such as using receptor-binding PDGF-D variants. Suitable antagonists include, but are not limited to, antibodies directed against PDGF-D; competitive or non-competitive inhibitors of binding of PDGF-D to the PDGF-D receptor(s), such as the receptor-binding or PDGF-D dimer-forming but non-mitogenic PDGF-D variants referred to above; and anti-sense nucleotide sequences as described below.

A method is provided for determining agents that bind to an activated truncated form of PDGF-D. The method comprises contacting an activated truncated form of PDGF-D with a test agent and monitoring binding by any suitable means. Agents can include both compounds and other proteins.

The invention provides a screening system for discovering agents that bind an activated truncated form of PDGF-D. The screening system comprises preparing an activated truncated form of PDGF-D, exposing the activated truncated form of PDGF-D to a test agent, and quantifying the binding of said agent to the

activated truncated form of PDGF-D by any suitable means. This screening system can also be used to identify agents which inhibit the proteolytic cleavage of the full length PDGF-D protein and thereby prevent the release of the activated truncated form of PDGF-D. For this use, the full length PDGF-D must be prepared.

Use of this screen system provides a means to determine compounds that may alter the biological function of PDGF-D. This screening method may be adapted to large-scale, automated procedures such as a PANDEX® (Baxter-Dade Diagnostics) system, allowing for efficient high-volume screening of potential therapeutic agents.

For this screening system, an activated truncated form of PDGF-D or full length PDGF-D is prepared as described herein, preferably using recombinant DNA technology. A test agent, e.g. a compound or protein, is introduced into a reaction vessel containing the activated truncated form of or full length PDGF-D. Binding of the test agent to the activated truncated form of or full length PDGF-D is determined by any suitable means which include, but is not limited to, radioactively- or chemically-labeling the test agent. Binding of the activated truncated form of or full length PDGF-D may also be carried out by a method disclosed in U.S. Patent 5,585,277, which is incorporated by reference. In this method, binding of the test agent to the activated truncated form of or full length PDGF-D is assessed by monitoring the ratio of folded protein to unfolded protein. Examples of this monitoring can include, but are not limited to, monitoring the sensitivity of the activated truncated form of or full length PDGF-D to a protease, or amenability to binding of the protein by a specific antibody against the folded state of the protein.

Those of skill in the art will recognize that IC_{50} values are dependent on the selectivity of the agent tested. For example, an agent with an IC_{50} which is less than 10 nM is generally

considered an excellent candidate for drug therapy. However, an agent which has a lower affinity, but is selective for a particular target, may be an even better candidate. Those skilled in the art will recognize that any information regarding the binding potential, inhibitory activity or selectivity of a particular agent is useful toward the development of pharmaceutical products.

Where PDGF-D or a PDGF-D antagonist is to be used for therapeutic purposes, the dose(s) and route of administration will depend upon the nature of the patient and condition to be treated, and will be at the discretion of the attending physician or veterinarian. Suitable routes include oral, subcutaneous, intramuscular, intraperitoneal or intravenous injection, parenteral, topical application, implants etc. Topical application of PDGF-D may be used in a manner analogous to VEGF. Where used for wound healing or other use in which enhanced angiogenesis is advantageous, an effective amount of the truncated active form of PDGF-D is administered to an organism in need thereof in a dose between about 0.1 and 1000 µg/kg body weight.

The PDGF-D or a PDGF-D antagonist may be employed in combination with a suitable pharmaceutical carrier. The resulting compositions comprise a therapeutically effective amount of PDGF-D or a PDGF-D antagonist, and a pharmaceutically acceptable non-toxic salt thereof, and a pharmaceutically acceptable solid or liquid carrier or adjuvant. Examples of such a carrier or adjuvant include, but are not limited to, saline, buffered saline, Ringer's solution, mineral oil, talc, corn starch, gelatin, lactose, sucrose, microcrystalline cellulose, kaolin, mannitol, dicalcium phosphate, sodium chloride, alginic acid, dextrose, water, glycerol, ethanol, thickeners, stabilizers, suspending agents and combinations thereof. Such compositions may be in the form of solutions, suspensions, tablets, capsules, creams, salves, elixirs, syrups, wafers, ointments or other conventional forms.

The formulation to suit the mode of administration. Compositions which comprise PDGF-D may optionally further comprise one or more of PDGF-A, PDGF-B, PDGF-C, VEGF, VEGF-B, VEGF-C, VEGF-D, PlGF and/or heparin. Compositions comprising PDGF-D will contain from
5 about 0.1% to 90% by weight of the active compound(s), and most generally from about 10% to 30%.

For intramuscular preparations, a sterile formulation, preferably a suitable soluble salt form of the truncated active form of PDGF-D, such as hydrochloride salt, can be dissolved and
10 administered in a pharmaceutical diluent such as pyrogen-free water (distilled), physiological saline or 5% glucose solution. A suitable insoluble form of the compound may be prepared and administered as a suspension in an aqueous base or a pharmaceutically acceptable oil base, e.g. an ester of a long
15 chain fatty acid such as ethyl oleate.

According to yet a further aspect, the invention provides diagnostic/prognostic devices typically in the form of test kits. For example, in one embodiment of the invention there is provided a diagnostic/prognostic test kit comprising an antibody to PDGF-D
20 and a means for detecting, and more preferably evaluating, binding between the antibody and PDGF-D. In one preferred embodiment of the diagnostic/prognostic device according to the invention, a second antibody (the secondary antibody) directed against antibodies of the same isotype and animal source of the antibody
25 directed against PDGF-D (the primary antibody) is provided. The secondary antibody is coupled directly or indirectly to a detectable label, and then either an unlabeled primary antibody or PDGF-D is substrate-bound so that the PDGF-D/primary antibody interaction can be established by determining the amount of label
30 bound to the substrate following binding between the primary antibody and PDGF-D and the subsequent binding of the labeled secondary antibody to the primary antibody. In a particularly preferred embodiment of the invention, the diagnostic/prognostic

device may be provided as a conventional enzyme-linked immunosorbent assay (ELISA) kit.

In another alternative embodiment, a diagnostic/prognostic device may comprise polymerase chain reaction means for
5 establishing sequence differences of a PDGF-D of a test individual and comparing this sequence structure with that disclosed in this application in order to detect any abnormalities, with a view to establishing whether any aberrations in PDGF-D expression are related to a given disease condition.

10 In addition, a diagnostic/prognostic device may comprise a restriction length polymorphism (RFLP) generating means utilizing restriction enzymes and genomic DNA from a test individual to generate a pattern of DNA bands on a gel and comparing this pattern with that disclosed in this application in order to detect
15 any abnormalities, with a view to establishing whether any aberrations in PDGF-D expression are related to a given disease condition.

In accordance with a further aspect, the invention relates to a method of detecting aberrations in PDGF-D gene structure in
20 a test subject which may be associated with a disease condition in the test subject. This method comprises providing a DNA sample from said test subject; contacting the DNA sample with a set of primers specific to PDGF-D DNA operatively coupled to a polymerase and selectively amplifying PDGF-D DNA from the sample by
25 polymerase chain reaction, and comparing the nucleotide sequence of the amplified PDGF-D DNA from the sample with the nucleotide sequences shown in Figure 3 (SEQ ID NO:3), Figure 5 (SEQ ID NO:5) or Figure 7 (SEQ ID NO:7). The invention also includes the provision of a test kit comprising a pair of primers specific to
30 PDGF-D DNA operatively coupled to a polymerase, whereby said polymerase is enabled to selectively amplify PDGF-D DNA from a DNA sample.

The invention also provides a method of detecting PDGF-D in a biological sample, comprising the step of contacting the sample with a reagent capable of binding PDGF-D, and detecting the binding. Preferably the reagent capable of binding PDGF-D is an antibody directed against PDGF-D, particularly a monoclonal antibody. In a preferred embodiment the binding and/or extent of binding is detected by means of a detectable label; suitable labels are discussed above.

In another aspect, the invention relates to a protein dimer comprising the PDGF-D polypeptide, particularly a disulfide-linked dimer. The protein dimers of the invention include both homodimers of PDGF-D polypeptide and heterodimers of PDGF-D and VEGF, VEGF-B, VEGF-C, VEGF-D, PlGF, PDGF-A, PDGF-B or PDGF-C.

According to a yet further aspect of the invention there is provided a method for isolation of PDGF-D comprising the step of exposing a cell which expresses PDGF-D to heparin to facilitate release of PDGF-D from the cell, and purifying the thus-released PDGF-D.

Another aspect of the invention involves providing a vector comprising an anti-sense nucleotide sequence which is complementary to at least a part of a DNA sequence which encodes PDGF-D or a fragment or analog thereof that has the biological activity of PDGF-D. In addition the anti-sense nucleotide sequence can be to the promoter region of the PDGF-D gene or other non-coding region of the gene which may be used to inhibit, or at least mitigate, PDGF-D expression.

According to a yet further aspect of the invention such a vector comprising an anti-sense sequence may be used to inhibit, or at least mitigate, PDGF-D expression. The use of a vector of this type to inhibit PDGF-D expression is favored in instances where PDGF-D expression is associated with a disease, for example where tumors produce PDGF-D in order to provide for angiogenesis, or tissue remodeling that takes place during invasion of tumor

cells into a normal cell population by primary or metastatic tumor formation. Transformation of such tumor cells with a vector containing an anti-sense nucleotide sequence would suppress or retard angiogenesis, and so would inhibit or retard growth of the tumor or tissue remodeling.

Another aspect of the invention relates to the discovery that the full length PDGF-D protein is likely to be a latent growth factor that needs to be activated by proteolytic processing to release an active PDGF/VEGF homology domain. A putative proteolytic site is found in residues 255-258 in the full length protein, residues -RKSK- (SEQ ID NO:9). This is a dibasic motif. The -RKSK- (SEQ ID NO:9) putative proteolytic site is also found in PDGF-A, PDGF-B, VEGF-C and VEGF-D. In these four proteins, the putative proteolytic site is also found just before the minimal domain for the PDGF/VEGF homology domain. Together these facts indicate that this is the proteolytic site.

Preferred proteases include, but are not limited, to plasmin, Factor X and enterokinase. The N-terminal CUB domain may function as an inhibitory domain which might be used to keep PDGF-D in a latent form in some extracellular compartment and which is removed by limited proteolysis when PDGF-D is needed.

According to this aspect of the invention, a method is provided for producing an activated truncated form of PDGF-D or for regulating receptor-binding specificity of PDGF-D. These methods comprise the steps of expressing an expression vector comprising a polynucleotide encoding a polypeptide having the biological activity of PDGF-D and supplying a proteolytic amount of at least one enzyme for processing the expressed polypeptide to generate the activated truncated form of PDGF-D.

This aspect also includes a method for selectively activating a polypeptide having a growth factor activity. This method comprises the step expressing an expression vector comprising a polynucleotide encoding a polypeptide having a growth factor

activity, a CUB domain and a proteolytic site between the polypeptide and the CUB domain, and supplying a proteolytic amount of at least one enzyme for processing the expressed polypeptide to generate the activated polypeptide having a growth factor activity.

In addition, this aspect includes the isolation of a nucleic acid molecule which codes for a polypeptide having the biological activity of PDGF-D and a polypeptide thereof which comprises a proteolytic site having the amino acid sequence RKSR (SEQ ID NO:9) or a structurally conserved amino acid sequence thereof.

Also this aspect includes an isolated dimer comprising an activated monomer of PDGF-D and an activated monomer of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-D, PDGF-A, PDGF-B, PDGF-C or PlGF linked to a CUB domain, or alternatively, an activated monomer of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-D, PDGF-A, PDGF-B or PlGF and an activated monomer of PDGF-D linked to a CUB domain. The isolated dimer may or may not include a proteolytic site between the activator monomer and the CUB domain linkage.

Polynucleotides of the invention such as those described above, fragments of those polynucleotides, and variants of those polynucleotides with sufficient similarity to the non-coding strand of those polynucleotides to hybridize thereto under stringent conditions all are useful for identifying, purifying, and isolating polynucleotides encoding other, non-human, mammalian forms of PDGF-D. Thus, such polynucleotide fragments and variants are intended as aspects of the invention. Exemplary stringent hybridization conditions are as follows: hybridization at 42°C in 5X SSC, 20 mM NaPO₄, pH 6.8, 50% formamide; and washing at 42°C in 0.2X SSC. Those skilled in the art understand that it is desirable to vary these conditions empirically based on the length and the GC nucleotide base content of the sequences to be hybridized, and that formulas for determining such variation exist. See for example Sambrook et al, "Molecular Cloning: A

Laboratory Manual", Second Edition, pages 9.47-9.51, Cold Spring Harbor, New York: Cold Spring Harbor Laboratory (1989).

Moreover, purified and isolated polynucleotides encoding other, non-human, mammalian PDGF-D forms also are aspects of the invention, as are the polypeptides encoded thereby and antibodies that are specifically immunoreactive with the non-human PDGF-D variants. Thus, the invention includes a purified and isolated mammalian PDGF-D polypeptide and also a purified and isolated polynucleotide encoding such a polypeptide.

It will be clearly understood that nucleic acids and polypeptides of the invention may be prepared by synthetic means or by recombinant means, or may be purified from natural sources.

It will be clearly understood that for the purposes of this specification the word "comprising" means "included but not limited to". The corresponding meaning applies to the word "comprises".

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 (SEQ ID NO:1) shows a nucleotide sequence that includes a cDNA sequence encoding the C-terminal part of human PDGF-D (hPDGF-D). The nucleotides which encode for the partial fragment of hPDGF-D are 1 to 198. The deduced partial amino acid sequence of hPDGF-D (66 amino acid residues-SEQ ID NO:2) derived from nucleotides 1 to 198 of Figure 1 is shown in Figure 2;

Figure 3 (SEQ ID NO:3) shows an extended sequence of a partial human cDNA encoding for the hPDGF-D. The translated cDNA sequence is from nucleotide 1 to 600. The deduced partial amino acid sequence of hPDGF-D (200 residues-SEQ ID NO:4) derived from nucleotides 1 to 600 of Figure 3 is shown in Figure 4;

Figure 5 shows a still further extended nucleotide sequence of a partial human cDNA. The nucleotides which encode for the 5'-truncated full-length hPDGF-D are 1 to 966 (SEQ ID NO:5). The deduced partial amino acid sequence of hPDGF-D (322 residues-SEQ

ID NO:6) derived from nucleotides 1 to 966 of Figure 5 is shown in Figure 6;

Figure 7 (SEQ ID NO:7) shows the complete nucleotide sequence of cDNA encoding a hPDGF-D(1116 bp) and the deduced amino acid sequence of full-length hPDGF-D encoded thereby which consists of 371 amino acid residues (Figure 8-SEQ ID NO:8);

Figure 9 shows an amino acid sequence alignment of the PDGF/VEGF-homology domain in hPDGF-D with several growth factors belonging to the VEGF/PDGF family (SEQ ID NOs:10-18, respectively);

Figure 10 shows a phylogenetic tree of several growth factors belonging to the VEGF/PDGF family;

Figure 11 provides the amino acid sequence alignment of the CUB domain present in hPDGF-D (SEQ ID NO:19) and other CUB domains present in human bone morphogenic protein-1 (hBMP-1, 3 CUB domains CUB1-3) (SEQ ID NOs:20-22, respectively) and in human neuropilin-1 (2 CUB domains) (SEQ ID NOs:23-24, respectively);

Figure 12 shows that conditioned medium(CM)containing truncated PDGF-D stimulates tyrosine phosphorylation of PDGF beta-receptors in PAE-1 cells; and

Figure 13 provides a graphical representation of results which show that conditioned medium(CM)containing truncated PDGF-D competes for binding with PDGF-BB homodimers for the PDGF beta-receptors, but not with PDGF-AA homodimers for the PDGF alpha-receptors.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

Figure 1 shows a nucleotide sequence of human cDNA which encodes a C-terminal portion of a novel growth factor, referred to herein as PDGF-D (formerly VEGF-G). PDGF-D is a new member of the VEGF/PDGF family. The nucleotide sequence of Figure 1 (SEQ ID NO:1) was derived from a human EST sequence (id. AI488780) in the dbEST database at the NCBI in Washington, DC. The nucleotides

1 to 198 of the cDNA of Figure 1 (SEQ ID NO:1) encodes a 66 amino acid polypeptide (Figure 2-SEQ ID NO:2) which shows some sequence similarity to the known members of the VEGF/PDGF family.

5 The amino acid sequence of the polypeptide encoded by the nucleotides 1 to 198 of the polynucleotide of Figure 1 (SEQ ID NO:1) is shown in Figure 2 (SEQ ID NO:2).

To generate more sequence information on human PDGF-D, a human fetal lung λ gt10 cDNA library was screened using a 327 bp polymerase chain reaction (PCR)-generated probe, based on the
10 originally identified EST sequence. The probe was generated from DNA from a commercially available human fetal lung cDNA library (Clontech) which was amplified by PCR using two primers derived from the identified EST (AI488780). The primers were:

5'-GTCGTGGAAGTGTCAACTGG (forward) (SEQ ID NO:26) and
15 5'-CTCAGCAACCACTTGTGTTC (reverse) (SEQ ID NO:27).

The amplified 327 bp fragment was cloned into the pCR2.1 vector (Invitrogen). Nucleotide sequencing verified that the insert corresponded to the EST. The screen identified several positive clones. The inserts from two of these clones, clones 5 and 8 were
20 subcloned into pBluescript and subjected to nucleotide sequencing using internal or vector-specific primers. The nucleotide sequences determined were identical in both clones and are shown in Figure 3 (SEQ ID NO:3). The coding region of the 690 bp polynucleotide is nucleotides 1-600 (SEQ ID NO:3) that encodes for
25 a large portion of hPDGF-D with the exception of the 5'-end. This portion of hPDGF-D includes the bioactive fragment of hPDGF-D. The deduced partial amino acid sequence of hPDGF-D (200 residues-SEQ ID NO:4) derived from nucleotides 1 to 600 of Figure 3 (SEQ ID NO:3) is shown in Figure 4 (SEQ ID NO:4).

30 Extended nucleotide sequencing of the isolated human PDGF-D cDNA clones from this human fetal lung cDNA library has provided additional sequence. Figure 5 (SEQ ID NO:5) shows a nucleotide sequence of a partial human cDNA (1934 bp) that encodes hPDGF-D.

The coding region of the 1934 bp polynucleotide is nucleotides 1 to 966 that encodes for hPDGF-D except for the most 5'-end of the polypeptide. The deduced partial amino acid sequence of hPDGF-D (322 residues-SEQ ID NO:6) derived from nucleotides 1 to 966 of Figure 5 (SEQ ID NO:5) is shown in Figure 6 (SEQ ID NO:6).

Figure 7 (SEQ ID NO:7) shows a polynucleotide sequence of cDNA encoding a full-length hPDGF-D. The region encoding PDGF-D is 1116 bp. The deduced amino acid sequence of full-length hPDGF-D is 371 amino acid residues (Figure 8-SEQ ID NO:8).

The sequence for the 5' end of full-length PDGF-D was obtained using Rapid Amplification of cDNA Ends (RACE) PCR, and clones containing cDNA from the human heart (Marathon-Ready™ cDNA, Clontech, Cat# 7404-1). These cDNA clones have an adaptor sequence attached to the 5' end of each clone, including a site for primer called Adaptor Primer 1 (Clontech: 5'9 CCATCCTAATACGACTCACTATAGGGC 3'9) (SEQ ID NO:28). This primer and a second primer 5'AGTGGGATCCGTTACTGA TGGAGAGTTAT 3' (SEQ ID NO:29) were used to amplify the sequence found at the 5' end of PDGF-D. In the PCR reaction a special polymerase mix was used (Advantage®-GC cDNA PCR Kit, Clontech, Cat# K1907-1). The reaction mix included (in microliters):

Adaptor Primer 1	1
Gene specific primer	1
Template (Human Heart cDNA)	5
GC-Melt (from the K1907-1 Kit)	5
5xGC cDNA PCR Reaction Buffer	10
50x dNTP mix	1
Sterile H2O	27
Total	50

The 5' end of PDGF-D was amplified for 31 cycles, five cycles consisted of 45 seconds denaturation at 94°C and four minutes extension at 72°C, five cycles consisted of 45 seconds

denaturation at 94°C and four minutes extension at 70°C, and five cycles consisted of 45 seconds denaturation at 94°C and four minutes extension at 68°C and an initial denaturation step at 94°C for two minutes. From this PCR, an approximately 790 bp long product was obtained. This product was run on a 1% agarose gel, purified (QIAquick gel extraction Kit, Qiagen, Cat # 28706) from the gel, cloned into a vector (TOPO TA Cloning Kit, Invitrogen) and transformed into bacteria (E.Coli). Transformed bacteria were plated, and incubated at 37°C overnight. Single colonies were picked and grown in fresh media overnight. Plasmids were prepared (QIAprep Spin Miniprep Kit, Qiagen, Cat# 27106) and sequenced with the plasmid primers, T7 and M13R. The result of this sequencing was that 312 bp of previously unknown PDGF-D sequence was obtained. The rest of the sequence (478 bp) was identical with previously obtained sequence from other PDGF-D cDNA clones.

Figure 9 shows the amino acid sequence alignment of the PDGF/VEGF-homology domain of PDGF-D (found in the C-terminal region of the polypeptide) with the PDGF/VEGF-homology domains of PDGF/VEGF family members, VEGF₁₆₅, PlGF-2, VEGF-B₁₆₇, Pox Orf VEGF, VEGF-C, VEGF-D, PDGF-A and PDGF-B (SEQ ID NOS:10-18, respectively). Some of the amino acid sequences in the N- and C-terminal regions in VEGF-C and VEGF-D have been deleted in this figure. Gaps were introduced to optimize the alignment. This alignment was generated using the MEGALIGN alignment tool based on the method of J. Hein, (Methods Enzymol. 1990 183 626-45) The PAM 250 residue weight table is used with a gap penalty of eleven and a gap length penalty of three and a K-tuple value of two in the pairwise alignments. The alignment is then refined manually, and the number of identities are estimated in the regions available for a comparison. The boxed residues indicate amino acids which match the VEGF-D within two distance units.

The alignment shows that PDGF-D has the expected pattern of invariant cysteine residues, a hallmark of members of this family,

with two exceptions. The first exception occurs between cysteine 3 and 4. Normally these two cysteines are spaced by 2 residues and with PDGF-D there is an insertion of three extra amino acids (NCA). This feature of the sequence in PDGF-D was highly unexpected. The second is that the invariant fifth cysteine found in the other members of the PDGF/VEGF family is not conserved in PDGF-D. This feature is unique to PDGF-D.

Based on the amino acid sequence alignments in Figure 9, a phylogenetic tree was constructed and is shown in Figure 10. The data show that the PVHD of PDGF-D is closely related to the PVHDs of VEGF-C and VEGF-D.

CUB Domain

The N-terminal region of the partial PDGF-D amino acid sequence of Figure 11 (residues 54-171) (SEQ ID NO:8) has second distinct protein domain which is referred to as a CUB domain (Bork and Beckmann, J. Mol. Biol., 1993 231, 539-545). This domain of about 115 amino acids was originally identified in complement factors C1r/C1s, but has recently been identified in several other extracellular proteins including signaling molecules such as bone morphogenic protein 1 (BMP-1) (Wozney et al., Science, 1988 242, 1528-1534) as well as in several receptor molecules such as neuropilin-1 (NP-1) (Soker et al., Cell, 1998 92 735-745). The functional roles of CUB domains are not clear but they may participate in protein-protein interactions or in interactions with carbohydrates including heparin sulfate proteoglycans. These interactions may play a role the proteolytic activation of PDGF-D.

As shown in Figure 11, the amino acid sequences from several CUB-containing proteins were aligned. The results show that the single CUB domain in human PDGF-D (SEQ ID NO:19) displays a significant identify with the most closely related CUB domains. Sequences from human BMP-1, with 3 CUB domains (CUBs1-3) (SEQ ID NOs:20-22, respectively) and human neuropilin-1 with 2 CUB domains

(CUBS1-2) (SEQ ID NOs:23-24, respectively) are shown. This alignment was generated as described above.

Example 1: Expression of PDGF-D transcripts

To investigate the tissue expression of PDGF-D in several human tissues, a Northern blot was done using a commercial Multiple Tissue Northern blot (MTN, Clontech). The blots were hybridized according to the instructions from the supplier using ExpressHyb solution at 68°C for one hour (high stringency conditions), and probed with the 327 bp PCR-generated probe from the human fetal lung cDNA library (see description above). The blots were subsequently washed at 50°C in 2X SSC with 0.05% SDS for 30 minutes and at 50°C in 0.1X SSC with 0.1% SDS for an additional 40 minutes. The blots were then put on film and exposed at -70°C. The results, summarized in Table 1, showed that expression of PDGF-D transcripts were most abundant in heart, pancreas and ovary while lower expression levels were seen in placenta, liver, kidney, prostate and testis. The human PDGF-D transcript was around 4 kb in length.

Table 1. Relative expression levels of PDGF-D transcripts in several human tissues as determined by Northern blot analysis

Tissue level*	Expression
Heart	+++++
Brain	n.d.
Placenta	++
Lung	+
Liver	++
Skeletal muscle	n.d.
Kidney	++
Pancreas	++++
Spleen	+
Thymus	+
Prostate	++
Testis	+++
Ovary	+++++

Small intestine ++
 Colon +
 Peripheral blood +
 leucocytes

5

The relative intensities of the bands were visually determined *
 (+++++) highest expression and (+) lowest expression; n.d. not
 detected

10

Example 2: Receptor binding properties of a truncated PDGF-D

To assess the interactions between a truncated PDGF-D and the
 VEGF receptors, truncated PDGF-D was tested for its capacity to
 bind to soluble Ig-fusion proteins containing the extracellular
 domains of human VEGFR-1, VEGFR-2 and VEGFR-3 (Olofsson et al.,
 Proc. Natl. Acad. Sci. USA, 1998 95 11709-11714). An expression
 vector encoding the PDGF/VEGF homology domain of PDGF-D was
 generated in the vector pSecTag (Invitrogen). The primers 5'-
 CCCAAGCTTGAAGATCTTGAGAATAT 3' (forward) (SEQ ID NO:30) and 5'-
 TGCTCTAGATCGAGGTGGTCTT 3' (reverse) (SEQ ID NO:31) were used to
 amplify a 429 bp fragment (nucleotides 556 to 966 in Figure 5) (SEQ
 ID NO:5) encoding amino acid residues 186 to 322 of Figure 6 (SEQ
 ID NO:6). The fragment was subsequently cloned into a HindIII and
 XbaI digested expression vector. COS cells were transfected with
 the expression vector encoding truncated PDGF-D or a control
 vector using calcium phosphate precipitation. The expressed
 polypeptide included a C-terminal c-myc tag and a 6X His tag (both
 derived from the pSecTag vector).

The Ig-fusion proteins, designated VEGFR-1-Ig, VEGFR-2-Ig and
 VEGFR-3-Ig, were transiently expressed in human 293 EBNA cells.
 All Ig-fusion proteins were human VEGFRs. Cells were incubated
 for 24 hours after transfection, washed with Dulbecco's Modified
 Eagle Medium (DMEM) containing 0.2% bovine serum albumin (BSA) and
 starved for 24 hours. The fusion proteins were then precipitated
 from the clarified conditioned medium using protein A-Sepharose
 beads (Pharmacia). The beads were combined with 100 microliters

of 10X binding buffer (5% BSA, 0.2% Tween 20 and 10 µg/ml heparin) and 900 microliter of conditioned medium prepared from the COS cells transfected with the expression vector for truncated PDGF-D or the control vector. The cells were then metabolically labeled with ³⁵S-cysteine and methionine (Promix, Amersham) for 4 to 6 hours. After 2.5 hours, at room temperature, the Sepharose beads were washed 3 times with binding buffer at 4°C, once with phosphate buffered saline (PBS) and boiled in SDS-PAGE buffer. Labeled proteins that were bound to the Ig-fusion proteins were analyzed by SDS-PAGE under reducing conditions. Radiolabeled proteins were detected using a phosphorimager analyzer and/or on film. In all these analyses, radiolabeled PDGF-D failed to show any interaction with any of the VEGF receptors. These results indicate that secreted truncated PDGF-D does not bind to VEGF receptors R1, R2 and R3.

Example 3: PDGF beta-receptor Phosphorylation

To test if PDGF-D causes increased phosphorylation of the PDGF beta-receptor, truncated PDGF-D was tested for its capacity to bind to the PDGF beta-receptor and stimulate increased phosphorylation. Serum-starved porcine aortic endothelial-1 (PAE-1) cells stably expressing the human PDGF beta-receptor (Eriksson et al., EMBO J, 1992, 11, 543-550) were incubated on ice for 90 minutes with a solution of conditioned media mixed with an equal volume of PBS supplemented with 1 mg/ml BSA. The conditioned media was prepared from COS cells transfected with expression vectors for PDGF-A or truncated PDGF-D (as constructed in Example 1), or a mock control vector. Twenty-four hours after transfection, the medium was replaced by serum-free medium containing 1mg/ml serum albumin. Conditioned medium was harvested after an additional 48 hours of incubation. Sixty minutes after the addition of the conditioned media, the cells were lysed in lysis buffer (20 mM tris-HCl, pH 7.5, 0.5% Triton X-100, 0.5%

deoxycholic acid, 10 mM EDTA, 1 mM orthovanadate, 1 mM PMSF 1% Trasylol). The PDGF beta-receptors were immunoprecipitated from cleared lysates with rabbit antisera against the human PDGF beta-receptor (Eriksson et al., EMBO J, 1992 11 543-550). The precipitated receptors were applied to a SDS-PAGE gel. After SDS gel electrophoresis, the precipitated receptors were transferred to nitrocellulose filters, and the filters were probed with anti-phosphotyrosine antibody PY-20, (Transduction Laboratories). The filters were then incubated with horseradish peroxidase-conjugated anti-mouse antibodies. Bound antibodies were detected using enhanced chemiluminescence (ECL, Amersham Inc). The filters were then stripped and reprobed with the PDGF beta-receptor rabbit antisera, and the amount of receptors was determined by incubation with horseradish peroxidase-conjugated anti-rabbit antibodies. Bound antibodies were detected using enhanced chemiluminescence (ECL, Amersham Inc). The probing of the filters with PDGF beta receptor antibodies confirmed that equal amounts of the receptor were present in all lanes. Human recombinant PDGF-BB (100ng/ml) and untreated cells were included in the experiment as a control. Figure 11 shows that truncated PDGF-D containing conditioned medium stimulated PDGF beta-receptor tyrosine phosphorylation. This indicates that truncated PDGF-D is a PDGF beta-receptor ligand/agonist.

Example 4: Competitive Binding Assay

Next, truncated PDGF-D was tested for its capacity to bind to the human PDGF beta-receptor by analyzing its ability to compete with PDGF-BB for binding to the PDGF beta-receptor. The binding experiments were performed on porcine aortic endothelial-1 (PAE-1) cells stably expressing the human PDGF alpha- and beta-receptors, respectively (Eriksson et al., EMBO J, 1992, 11, 543-550). Binding experiments were performed essentially as in Heldin et al. (EMBO J, 1988, 7 1387-1393). Conditioned media from COS

cells expressing PDGF-A, truncated PDGF-D, or mock control, respectively, was diluted with an equal volume of BSA/PBS and mixed with 100 ng/ml of ^{125}I -PDGF-BB (beta-receptor ligand) or of ^{125}I -PDGF-AA (alpha-receptor ligand) in binding buffer (PBS containing 1 mg/ml of BSA). Two separate sets of conditioned media from these COS cells were analyzed. Aliquots were incubated with the receptor expressing PAE-1 cells plated in 24-well culture dishes on ice for 90 minutes. After three washes with binding buffer, cell-bound ^{125}I -PDGF-BB or ^{125}I -PDGF-AA was extracted by lysis of cells in 20 mM Tris-HCl, pH 7.5, 10% glycerol, 1% Triton X-100. The amount of cell bound radioactivity was determined in a gamma-counter. Figure 12 provides a graphical representation of results which show that conditioned medium containing truncated PDGF-D competes for binding with PDGF-BB homodimers for the PDGF beta-receptors, but not with PDGF-AA homodimers for the PDGF alpha-receptors.

PDGF-D does not bind to any of the known VEGF receptors. PDGF-D is the only VEGF family member, thus far, which can bind to and increase phosphorylation of the PDGF beta-receptor. These characteristics indicate that the truncated form of PDGF-D may not be a VEGF family member, but instead a novel PDGF. Furthermore, the full length protein is likely to be a latent growth factor that needs to be activated by proteolytic processing to release the active PDGF/VEGF homology domain. The N-terminal CUB domain may be expressed as an inhibitory domain which might be used to localize this latent growth factor in some extracellular compartment (for example the extracellular matrix) and which is removed by limited proteolysis when need, for example during embryonic development, tissue regeneration, tissue remodelling including bone remodelling, active angiogenesis, tumor progression, tumor invasion, metastasis formation and/or wound healing.

BIOASSAYS TO DETERMINE THE FUNCTION OF PDGF-D

Assays are conducted to evaluate whether PDGF-D has similar activities to PDGF-A, PDGF-B, VEGF, VEGF-B, VEGF-C and/or VEGF-D in relation to growth and/or motility of connective tissue cells, fibroblasts, myofibroblasts and glial cells; to endothelial cell function; to angiogenesis; and to wound healing. Further assays may also be performed, depending on the results of receptor binding distribution studies.

I. Mitogenicity of PDGF-D for Endothelial Cells

To test the mitogenic capacity of PDGF-D for endothelial cells, the PDGF-D polypeptide is introduced into cell culture medium containing 5% serum and applied to bovine aortic endothelial cells (BAEs) propagated in medium containing 10% serum. The BAEs are previously seeded in 24-well dishes at a density of 10,000 cells per well the day before addition of the PDGF-D. Three days after addition of this polypeptide the cells were dissociated with trypsin and counted. Purified VEGF is included in the experiment as positive control.

II. Mitogenicity of PDGF-D for Fibroblasts

To test the mitogenic capacity of PDGF-D for fibroblasts, different concentrations of truncated homodimers of PDGF-DD or PDGF-AA (as control) are added to serum starved human foreskin fibroblasts in the presence of 0.2 μ M [3H]thymidine. The fibroblasts are then incubated for 24 hours with 1 ml of serum-free medium supplemented with 1 mg/ml BSA. After trichloroacetic acid (TCA) precipitation, the incorporation of [3H]thymidine into DNA is determined using a beta-counter. The assay is performed essentially as described in Mori et al., J. Biol. Chem., 1991 266 21158-21164.

III. Assays of Endothelial Cell Function

a) *Endothelial cell proliferation*

Endothelial cell growth assays are performed by methods well known in the art, e.g. those of Ferrara & Henzel, *Nature*, 1989 380 439-443, Gospodarowicz et al., *Proc. Natl. Acad. Sci. USA*, 1989 86 7311-7315, and/or Claffey et al., *Biochem. Biophys. Acta*, 1995 1246 1-9.

b) *Cell adhesion assay*

The effect of PDGF-D on adhesion of polymorphonuclear granulocytes to endothelial cells is tested.

c) *Chemotaxis*

The standard Boyden chamber chemotaxis assay is used to test the effect of PDGF-D on chemotaxis.

d) *Plasminogen activator assay*

Endothelial cells are tested for the effect of PDGF-D on plasminogen activator and plasminogen activator inhibitor production, using the method of Pepper et al., *Biochem. Biophys. Res. Commun.*, 1991 181 902-906.

e) *Endothelial cell Migration assay*

The ability of PDGF-D to stimulate endothelial cells to migrate and form tubes is assayed as described in Montesano et al., *Proc. Natl. Acad. Sci. USA*, 1986 83 7297-7301. Alternatively, the three-dimensional collagen gel assay described in Joukov et al., *EMBO J.*, 1996 15 290-298 or a gelatinized membrane in a modified Boyden chamber (Glaser et al., *Nature*, 1980 288 483-484) may be used.

IV. Angiogenesis Assay

The ability of PDGF-D to induce an angiogenic response in chick chorioallantoic membrane is tested as described in Leung et al., Science, 1989 246 1306-1309. Alternatively the rat cornea assay of Rastinejad et al., Cell, 1989 56 345-355 may be used; this is an accepted method for assay of *in vivo* angiogenesis, and the results are readily transferrable to other *in vivo* systems.

V. Wound Healing

The ability of PDGF-D to stimulate wound healing is tested in the most clinically relevant model available, as described in Schilling et al., Surgery, 1959 46 702-710 and utilized by Hunt et al., Surgery, 1967 114 302-307.

VI. The Haemopoietic System

A variety of *in vitro* and *in vivo* assays using specific cell populations of the haemopoietic system are known in the art, and are outlined below. In particular a variety of *in vitro* murine stem cell assays using fluorescence-activated cell sorter to purified cells are particularly convenient:

a) Repopulating Stem Cells

These are cells capable of repopulating the bone marrow of lethally irradiated mice, and have the Lin⁻, Rh^{h1}, Ly-6A/E⁺, c-kit⁺ phenotype. PDGF-D is tested on these cells either alone, or by co-incubation with other factors, followed by measurement of cellular proliferation by ³H-thymidine incorporation.

b) Late Stage Stem Cells

These are cells that have comparatively little bone marrow repopulating ability, but can generate D13 CFU-S. These cells have the Lin⁻, Rh^{h1}, Ly-6A/E⁺, c-kit⁺ phenotype. PDGF-D is incubated with these cells for a period of time, injected into

lethally irradiated recipients, and the number of D13 spleen colonies enumerated.

c) *Progenitor-Enriched Cells*

5 These are cells that respond *in vitro* to single growth factors and have the Lin⁻, Rh^{h1}, Ly-6A/E⁺, c-kit⁺ phenotype. This assay will show if PDGF-D can act directly on haemopoietic progenitor cells. PDGF-D is incubated with
10 these cells in agar cultures, and the number of colonies present after 7-14 days is counted.

VII. Atherosclerosis

Smooth muscle cells play a crucial role in the development or initiation of atherosclerosis, requiring a change of their
15 phenotype from a contractile to a synthetic state. Macrophages, endothelial cells, T lymphocytes and platelets all play a role in the development of atherosclerotic plaques by influencing the growth and phenotypic modulations of smooth muscle cell. An *in vitro* assay using a modified Rose chamber in which different cell
20 types are seeded on to opposite cover slips measures the proliferative rate and phenotypic modulations of smooth muscle cells in a multicellular environment, and is used to assess the effect of PDGF-D on smooth muscle cells.

25 **VIII. Metastasis**

The ability of PDGF-D to inhibit metastasis is assayed using the Lewis lung carcinoma model, for example using the method of Cao et al., J. Exp. Med., 1995 182 2069-2077.

30 **IX. Migration of Smooth Muscle Cells**

The effects of the PDGF-D on the migration of smooth muscle cells and other cells types can be assayed using the method of Koyama et al., J. Biol. Chem., 1992 267 22806-22812.

X. Chemotaxis

The effects of the PDGF-D on chemotaxis of fibroblast, monocytes, granulocytes and other cells can be assayed using the method of Siegbahn et al., J. Clin. Invest., 1990 85 916-920.

XI. PDGF-D in Other Cell Types

The effects of PDGF-D on proliferation, differentiation and function of other cell types, such as liver cells, cardiac muscle and other cells, endocrine cells and osteoblasts can readily be assayed by methods known in the art, such as ³H-thymidine uptake by *in vitro* cultures.

XII. Construction of PDGF-D Variants and Analogues

PDGF-D is a member of the PDGF family of growth factors which exhibits a high degree of homology to the other members of the PDGF family. PDGF-D contains seven conserved cysteine residues which are characteristic of this family of growth factors. These conserved cysteine residues form intra-chain disulfide bonds which produce the cysteine knot structure, and inter-chain disulfide bonds that form the protein dimers which are characteristic of members of the PDGF family of growth factors. PDGF-D interacts with a protein tyrosine kinase growth factor receptor.

In contrast to proteins where little or nothing is known about the protein structure and active sites needed for receptor binding and consequent activity, the design of active mutants of PDGF-D is greatly facilitated by the fact that a great deal is known about the active sites and important amino acids of the members of the PDGF family of growth factors.

Published articles elucidating the structure/activity relationships of members of the PDGF family of growth factors include for PDGF: Oestman et al., J. Biol. Chem., 1991 266 10073-10077; Andersson et al., J. Biol. Chem., 1992 267 11260-1266; Oefner et al., EMBO J., 1992 11 3921-3926; Flemming et al.,

Molecular and Cell Biol., 1993 13 4066-4076 and Andersson et al., Growth Factors, 1995 12 159-164; and for VEGF: Kim et al., Growth Factors, 1992 7 53-64; Pötgens et al., J. Biol. Chem., 1994 269 32879-32885 and Claffey et al., Biochem. Biophys. Acta, 1995 1246 1-9. From these publications it is apparent that because of the eight conserved cysteine residues, the members of the PDGF family of growth factors exhibit a characteristic knotted folding structure and dimerization, which result in formation of three exposed loop regions at each end of the dimerized molecule, at which the active receptor binding sites can be expected to be located.

Based on this information, a person skilled in the biotechnology arts can design PDGF-D mutants with a very high probability of retaining PDGF-D activity by conserving the eight cysteine residues responsible for the knotted folding arrangement and for dimerization, and also by conserving, or making only conservative amino acid substitutions in the likely receptor sequences in the loop 1, loop 2 and loop 3 region of the protein structure.

The formation of desired mutations at specifically targeted sites in a protein structure is considered to be a standard technique in the arsenal of the protein chemist (Kunkel et al., Methods in Enzymol., 1987 154 367-382). Examples of such site-directed mutagenesis with VEGF can be found in Pötgens et al., J. Biol. Chem., 1994 269 32879-32885 and Claffey et al., Biochem. Biophys. Acta, 1995 1246 1-9. Indeed, site-directed mutagenesis is so common that kits are commercially available to facilitate such procedures (e.g. Promega 1994-1995 Catalog., Pages 142-145).

The connective tissue cell, fibroblast, myofibroblast and glial cell growth and/or motility activity, the endothelial cell proliferation activity, the angiogenesis activity and/or the wound healing activity of PDGF-D mutants can be readily confirmed by well established screening procedures. For example, a procedure

analogous to the endothelial cell mitotic assay described by Claffey, et al., (Biochem. Biophys. Acta., 1995 1246 1-9) can be used. Similarly the effects of PDGF-D on proliferation of other cell types, on cellular differentiation and on human metastasis can be tested using methods which are well known in the art.

5 The foregoing description and examples have been set forth merely to illustrate the invention and are not intended to be limiting. Since modifications of the disclosed embodiments incorporating the spirit and substance of the invention may occur
10 to persons skilled in the art, the invention should be construed broadly to include all variations falling within the scope of the appended claims and equivalents thereof.

What is claimed is:

1. An isolated nucleic acid molecule comprising a polynucleotide sequence having at least 85% identity with at least nucleotides 1 to 600 of the sequence set out in Figure 3 (SEQ ID NO:3), at least nucleotides 1 to 966 of the sequence set out in Figure 5 (SEQ ID NO:5), at least nucleotides 176-1288 of Figure 7 (SEQ ID NO:5) or at least nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7).
2. An isolated nucleic acid molecule according to claim 1, wherein the sequence identity is at least 90%.
3. An isolated nucleic acid molecule according to claim 1, wherein the sequence identity is at least 95%.
4. An isolated nucleic acid molecule according to claim 1, wherein said nucleic acid is a cDNA.
5. An isolated nucleic acid molecule according to claim 1, wherein said nucleic acid is a mammalian polynucleotide.
6. An isolated nucleic acid molecule according to claim 5, wherein said nucleic acid is a human polynucleotide.
7. An isolated nucleic acid molecule which encodes a polypeptide molecule comprising an amino acid sequence having at least 85% identity with the amino acid sequence of Figure 4 (SEQ ID NO:4), the amino acid sequence of Figure 6 (SEQ ID NO:6), or the amino acid sequence of Figure 8 (SEQ ID NO:8), or a fragment or analog thereof having the biological activity of PDGF-D.

8. An isolated nucleic acid molecule according to claim 7, wherein the amino acid sequence identity is at least 90%.
9. An isolated nucleic acid molecule according to claim 7, wherein the amino acid sequence identity is at least 95%.
10. An isolated nucleic acid molecule which codes for a polypeptide which comprises the amino acid sequence
PXCLLVXRCGGNCXC (SEQ ID NO:25).
11. An isolated polynucleotide, comprising a polynucleotide sequence having at least 85% identity with at least nucleotides 1 to 600 of the sequence set out in Figure 3 (SEQ ID NO:3), at least nucleotides 1 to 966 of the sequence set out in Figure 5 (SEQ ID NO:5), at least nucleotides 176-1288 of Figure 7 (SEQ ID NO:5) or at least nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7), or a polynucleotide which hybridizes under stringent conditions with at least one of said DNA sequences.
12. A vector comprising a nucleic acid according to claim 1 or claim 11, which nucleic acid is operably linked with a promoter sequence.
13. A vector according to claim 12, wherein said vector is a eukaryotic vector.
14. A vector according to claim 12, wherein said vector is a prokaryotic vector.
15. A vector according to claim 12, wherein said vector is a plasmid.

16. A vector according to claim 12, wherein said vector is a baculovirus vector.
17. A method of making a vector which expresses a polypeptide comprising an amino acid sequence having at least 85% identity with the amino acid sequence of Figure 4 (SEQ ID NO:4), the amino acid sequence of Figure 6 (SEQ ID NO:6), or the amino acid sequence of Figure 8 (SEQ ID NO:8), or fragment or analog thereof having the biological activity of PDGF-D, said method comprising incorporating an isolated nucleic acid according to claim 1, claim 7, claim 10 or claim 11, into said vector in operatively linked relation with a promoter.
18. A host cell transformed or transfected with a vector according to claim 12.
19. A host cell according to claim 18, wherein said host cell is a eukaryotic cell.
20. A host cell according to claim 18, wherein said host cell is a COS cell.
21. A host cell according to claim 18, wherein said host cell is a prokaryotic cell.
22. A host cell according to claim 18, wherein said host cell is a 293EBNA cell.
23. A host cell according to claim 18, wherein said host cell is an insect cell.
24. A host cell transformed or transfected with a vector comprising a nucleic acid sequence according to claim 1 or claim

11, operatively linked to a promoter, such that said host cell expresses a polypeptide comprising an amino acid sequence having at least 85% identity with the amino acid sequence of Figure 4 (SEQ ID NO:4), the amino acid sequence of Figure 6 (SEQ ID NO:6), or the amino acid sequence of Figure 8 (SEQ ID NO:8), or a fragment or analog thereof having the biological activity of PDGF-D.

25. An isolated polypeptide comprising at least 85% identity with the amino acid sequence of Figure 4 (SEQ ID NO:4), the amino acid sequence of Figure 6 (SEQ ID NO:6), or the amino acid sequence of Figure 8 (SEQ ID NO:8), or a fragment or analog thereof having the biological activity of PDGF-D.

26. An isolated polypeptide according to claim 25, wherein said polypeptide is a human polypeptide.

27. An isolated polypeptide according to claim 25, wherein said polypeptide has the ability to stimulate and/or enhance proliferation and/or differentiation and/or growth and/or motility of cells expressing a PDGF-D receptor.

28. An isolated polypeptide according to claim 27, wherein the cells are selected from the group consisting of endothelial cells, connective tissue cells, myofibroblasts and glial cells.

29. An isolated polypeptide produced by expression of a polynucleotide comprising the polynucleotide sequence having at least 85% identity with at least nucleotides 1 to 600 of the sequence set out in Figure 3 (SEQ ID NO:3), at least nucleotides 1 to 966 of the sequence set out in Figure 5 (SEQ ID NO:5), at least nucleotides 176-1288 of Figure 7 (SEQ ID NO:5) or at least nucleotides 938 to 1288 set out in Figure 7 (SEQ ID NO:7), or a

polynucleotide which hybridizes under stringent conditions with at least one of said DNA sequences.

30. An isolated polypeptide which comprises the characteristic sequence

PXCLLVXRCGGNCXC (SEQ ID NO:25).

31. An isolated polypeptide dimer comprising a polypeptide according to claim 25 or claim 29.

32. An isolated polypeptide dimer according to claim 31, wherein said polypeptide dimer is a homodimer of said polypeptide.

33. An isolated polypeptide dimer according to claim 31, wherein said polypeptide dimer is a heterodimer of said polypeptide and VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-A, PDGF-B or PlGF.

34. An isolated polypeptide dimer according to claim 31, wherein said polypeptide dimer is a disulfide-linked dimer.

35. A pharmaceutical composition comprising an effective cell proliferation promoting amount of a polypeptide according to claim 25, claim 29 or claim 30, and at least one further growth factor selected from the group consisting of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-A, PDGF-B or PlGF.

36. A pharmaceutical composition according to claim 35, further comprising heparin.

37. A pharmaceutical composition comprising an effective cell proliferation promoting amount of an isolated polypeptide

according to claim 25, claim 29 or claim 30, and at least one pharmaceutical carrier or diluent.

38. A pharmaceutical composition according to claim 37, further comprising heparin.

39. A pharmaceutical composition comprising an effective amount of an isolated polypeptide according to claim 25, claim 29 or claim 30 and heparin.

40. A pharmaceutical composition comprising a PDGF receptor stimulating amount of an isolated polypeptide according to claim 25, claim 29 or claim 30, and at least one pharmaceutical carrier or diluent.

41. A means for amplifying a polynucleotide according to claim 1 or claim 11 in a test sample, said means comprising at least one pair of primers complementary to a nucleic acid according to claim 1 or claim 11.

42. A means for amplifying a polynucleotide as according to claim 1 or claim 11 in a test sample, said means comprising a polymerase and at least one pair of primers complementary to a nucleic acid according to claim 1 or claim 11, for amplifying the polynucleotide by polymerase chain reaction in order to facilitate a sequence comparison of the polynucleotide with the nucleic acid according to claim 1 or claim 11.

43. An antibody specifically reactive with a polypeptide according to claim 25, claim 29 or claim 30.

44. An antibody according to claim 43, wherein said antibody is a polyclonal antibody.

45. An antibody according to claim 43, wherein said antibody is a monoclonal antibody.

46. An antibody according to claim 45, wherein said antibody is a humanized antibody.

47. An antibody according to claim 44 or 45 or 46, wherein said antibody is labeled with a detectable label.

48. An antibody according to claim 47, wherein said detectable label is radioactive isotope.

49. An antibody according to claim 45 or 46, wherein the antibody is modified by addition of cytotoxic or cytostatic drug.

50. A method of making a polypeptide according to claim 25, claim 29 or claim 30, said method comprising the steps of:
culturing a host cell transformed or transfected with a vector comprising a polynucleotide encoding said polypeptide operably associated with a promoter sequence such that the nucleic acid sequence encoding said polypeptide is expressed; and
isolating said polypeptide from said host cell or from a growth medium in which said host cell is cultured.

51. A method of stimulating growth of connective tissue or wound healing in a mammal comprising the step of administering to said mammal an effective connective tissue or wound healing stimulating amount of the polypeptide according to claim 25, claim 29 or 30.

52. A method for producing an activated truncated form of PDGF-D, comprising the steps of expressing an expression vector comprising a polynucleotide encoding a polypeptide as claimed in

claim 25, claim 29 or claim 30 and supplying a proteolytic amount of at least one enzyme for processing the expressed polypeptide to generate the activated truncated form of PDGF-D.

53. A method for regulating receptor-binding specificity of PDGF-D, comprising the steps of expressing an expression vector comprising a polynucleotide encoding a polypeptide as claimed in claim 25, claim 29 or claim 30 and supplying a proteolytic amount of at least one enzyme for processing the expressed polypeptide to generate the activated truncated form of PDGF-D.

54. A method for selectively activating a polypeptide having a growth factor activity comprising the step expressing an expression vector comprising a polynucleotide encoding a polypeptide having a growth factor activity, a CUB domain and a proteolytic site between the polypeptide and the CUB domain, and supplying a proteolytic amount of at least one enzyme for processing the expressed polypeptide to generate the activated polypeptide having a growth factor activity.

55. An isolated polypeptide according to claim 25, claim 29 or claim 30 which comprises a proteolytic site having the amino acid sequence RKSK or a structurally conserved amino acid sequence thereof.

56. An isolated nucleic acid molecule according to claim 7 which codes for a polypeptide which comprises a proteolytic site having the amino acid sequence RKSK or a structurally conserved amino acid sequence thereof.

57. An isolated heterodimer comprising an activated monomer of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-D, PDGF-A, PDGF-B or PlGF and an activated monomer of PDGF-D linked to a CUB domain.

58. An isolated heterodimer comprising an activated monomer of PDGF-D and an activated monomer of VEGF, VEGF-B, VEGF-C, VEGF-D, PDGF-D, PDGF-A, PDGF-B or PlGF linked to a CUB domain.

59. An isolated heterodimer according to claim 57, further comprising a proteolytic site between the activator monomer and the CUB domain linkage.

60. An isolated heterodimer according to claim 58, further comprising a proteolytic site between the activator monomer and the CUB domain linkage.

61. A method of inducing PDGF beta-receptor activation, comprising the step of adding a PDGF beta-receptor stimulating amount of the polypeptide according to claim 25, claim 29 or claim 30.

62. A method of inhibiting tumor growth of a tumor expressing PDGF-D in a mammal, comprising the step of administering to said mammal a PDGF-D inhibiting amount of a PDGF-D antagonist.

63. A method of identifying specific types of human tumors, comprising the step of taking a sample of the tumor and testing for the expression of PDGF-D.

64. A method for identifying an PDGF-D antagonist comprising:

admixing a substantially purified preparation of an activated truncated form of PDGF-D with a test agent; and
monitoring, by any suitable means, an inhibition in the biological activity of PDGF-D.

65. A method for identifying an PDGF-D antagonist comprising:

admixing a substantially purified preparation of an full-length PDGF-D with a test agent; and

monitoring, by any suitable means, an inhibition in the cleavage of the CUB domain from PDGF-D.

66. A method of making a vector which expresses a polypeptide comprising an amino acid sequence having at least 85% identity with the amino acid residues 255 to 371 of Figure 8 (SEQ ID NO:8), said method comprising incorporating an isolated nucleic acid molecule encoding said amino acid residues into said vector in operatively linked relation with a promoter.

67. A method for producing an activated truncated form of PDGF-D, comprising the steps of expressing an expression vector comprising a polynucleotide encoding a polypeptide as claimed in claim 25, claim 29 or claim 30 and supplying a proteolytic amount of at least one enzyme for processing the expressed polypeptide to generate the activated truncated form of PDGF-D.

68. A method of inhibiting tissue remodeling during invasion of tumor cells into a normal population of cells, comprising administering to said mammal a PDGF-D inhibiting amount of a PDGF-C antagonist.

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aattgtggct gtggaactgt caactggagg tcttgccat gcaattcagg gaaaaccgtg 60
 aaaaagttatc atgaggtatt acagtttgag cctggccaca tcaagaggag gggtagagct 120
 aagaccatgg ctctagttga catccagttg gatcaccatg aacgatgtga ttgtatctgc 180
 agtcaagac caccicgata agagaatgig cacatcctta catlaagcct gaaagaacca 240
 ttagtttaag gagggigaga taagagaccc ttttccctacc agcaaccaga cttactacta 300
 gccctgcaatg caatgaacac aagtggttgc tgagttctcat ccttgctttg ttaatgccat 360

FIG.1

Asn Cys Gly Cys Gly Thr Val Asn Trp Arg Ser Cys Thr Cys Asn Ser
 1 5 10 15
 Gly Lus Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro Gly
 20 25 30
 His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile
 35 40 45
 Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 50 55 60
 Pro Arg
 65

FIG.2

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ggaagatttc caaccgcag cagcttcaga gaccaactgg aatctgtcac aagctctgtt 60
tcagggatc cctataactc tccatcagta acggatccca ctctgattgc ggaatgctctg 120
gacaaaaaaa ttgcagaatt tgatcagtg gaagatctgc tcaagtactt caatccagag 180
tcatggcaag aafatcttga gaatatgtat ctggacaccc ctcggtatcg aggcaggtea 240
taccatgacc ggaagtcaaa agttgacctg gataggetca atgatgatgc caagcgltac 300
agttgcactc ccaggaatta ctcggtcaat alaagagaag agctgaagtt ggccaatgtg 360
gtcttctttc cacgttgccct cctcgtgcag cgctgtggag gaaattgttg ctgtggaact 420
gtcaaactgg agtccgtcac atgcaattca gggaaaaccg tgaaaaagta tcatgaggta 480
ttacagtttg agcctggcca catcaagagg aggggtagag ctaagaccat ggcctagtt 540
gacatccagt tggatcacca tgaacgatgc gatgtatct gcagctcaag accacctcga 600
taagagaatg tgcacatcct tacattaagc ctgaagaac ctltagttta aggaggggtga 660
gataagagac ccttttcta ccagcaacc 690

FIG.3

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Gly Arg Phe Pro Thr Arg Ser Ser Phe Arg Asp Gln Leu Glu Ser Val
 1 5 10 15
 Thr Ser Ser Val Ser Gly Tyr Pro Tyr Asn Ser Pro Ser Val Thr Asp
 20 25 30
 Pro Thr Leu Ile Ala Asp Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp
 35 40 45
 Thr Val Glu Asp Leu Leu Lys Tyr Phe Asn Pro Glu Ser Trp Gln Glu
 50 55 60
 Asp Leu Glu Asn Met Tyr Leu Asp Thr Pro Arg Tyr Arg Gly Arg Ser
 65 70 75 80
 Tyr His Asp Arg Lys Ser Lys Val Asl Leu Asp Arg Leu Asn Asp Asp
 85 90 95
 Ala Lys Arg Tyr Ser Cys Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg
 100 105 110
 Glu Glu Leu Lys Leu Ala Asn Val Val Phe Phe Pro Arg Cys Leu Leu
 115 120 125
 Val Gln Arg Cys Gly Gly Asn Cys Gly Cys Gly Thr Val Lys Leu Glu
 130 135 140
 Ser Cys Thr Cys Asn Ser Gly Lys Thr Val Lys Lus Tyr His Glu Val
 145 150 155 160
 Leu Gln Phe Glu Pro Gly His Ile Lys Arg Arg Gly Arg Ala Lys Thr
 165 170 175
 Met Ala Leu Val Asp Ile Gln Leu Asp His His Glu Arg Cys Asp Cys
 180 185 190
 Ile Cys Ser Ser Arg Pro Pro Arg
 195 200

FIG.4

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ttgtaccgaa gagatgagac catccagglg aaaggoaacg gctacgtlga gaglccclaga 60
 tccccgaaca gctaccccag gaacctgclt clgacatggc ggcttcactc lcaggagaat 120
 acacggalac agctagtgtt lgacaalcag tllggattag aggaagcaga aatlgatatt 180
 tglaggtatg allltltgga agltgaogat alalcgaaa ccagtlacat lallagagga 240
 cgalggltg gacacaagga agltccclca aggalaaaat caagaacgaa ccaaaltaaa 300
 atcacallca agtccgatlga clactlltltg gclaaacctg gatlcaogat llatlaltct 360
 ttgctggaa aglttcaacc cgcagcagct lcagagacca actgggaatc tglcacaagc 420
 tctatllcag ggglatclla laactclcca lcaglaacgg alccacrcr gattlclfat 480
 gctclggaca aaaaaatlge agaatltgat acaglggaag alclgclcaa glacttcaat 540
 ccagaglcac ggcaagaaga tcltgagaat atglalclgg acacccctcg gatltagggc 600
 agglcatacc atgaccgga gtcaaaaglt gacctggala ggclcaatga lgalgccaag 660
 cgttacaglt gcactccag gaattacteg glcaatataa gagaagagcl gaagltggcc 720
 aatgtggct tclttccag ttgcctclt glcagcgccl glggaggaaa lltgtggctgt 780
 ggaactglca actggagglt clgcacatgc aaltcagggg aaacctlga aaaglatcat 840
 gaggtaltac aglttgagcc tggccacalc aagaggaggg glagagclaa gaccatggct 900
 clagltlga tccagltgga lcacatgaa cgalgcgalt glatclgcag clcaagacca 960
 cclcgataag agaatglga catcltaca tlaagcclga aagaacctt aglttlaagga 1020
 gggtagata agagacctt lccclaccag caaccaact tactactagc clgcaatgca 1080
 algaacacaa gtggltgclg agtclcagcc lltclltgtt aatgccatgg caagtagaaa 1140
 gglatalcat caacttclal acctoagaat alaggatlgc attlaataal agltltlgag 1200
 gltatlatg cacaacaca cacagaaala laltcalgtc latgtgtata tagatcaaat 1260
 gllllllllt glatatataa ccaggtacac cagagcltac alalgtltga gtlagactct 1320
 laaaatclt lgcacaaata agggatgglt aatlatatga aacatglclt tagaaaatt 1380
 aggagataaa lltatlllt aattltgaaa cacaacacaa lltlgaaatc tgcctcltla 1440
 aagaagcat cltgtatatt aaaaatcaaa agatgaggcl lcltlacala tacatcltag 1500
 ttgattatta aaaaaggaaa aaggtltcca gagaaaaggc caatacc taa gcattlltcl 1560
 catgagaagc actgcalact tacctatgtg gactgtata acctgtclcc aaacacatgc 1620
 cataataala taagtcltlt agaaatltaa tcaatgtgt lltlatgcat lltgtlgagg 1680
 catcctatt catllaacac clatclcaaa aacttactta gaaggltllt laltataglc 1740
 clacaaaaga caatglataa gclglaacag aatlttgaat lgtlltclt lgcacaaacc 1800
 clccacaaaa gcaaatclt lcaagaatgg calggacat clglatgaac cltccagat 1860
 ggtgttcagl gaagatgtg gglagltlgag aacttaaaaa gtgaacallg aaacatcgac 1920
 glaactlgaa accg 1934

FIG.5

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Leu Tyr Arg Arg Asp Glu Thr Ile Gln Val Lys Gly Asn Gly Tyr Val
 1 5 10 15
 Gln Ser Pro Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Leu Thr
 20 25 30
 Trp Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp
 35 40 45
 Asn Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp
 50 55 60
 Phe Val Glu Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg Gly
 65 70 75 80
 Arg Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr
 85 90 95
 Asn Gln Ile Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala Lys
 100 105 110
 Pro Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro Ala
 115 120 125
 Ala Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser Gly
 130 135 140
 Val Ser Try Asn Ser Pro Ser Val The Asp Pro Thr Lei Ile Ala Asp
 145 150 155 160
 Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp The Val Glu Asp Leu Leu
 165 170 175
 Lys Tyr Phe Asn Pro Glu Ser Trp Gln Glu Asp Leu Glu Asn Met Tyr
 180 185 190
 Leu Asp Thr Pro Arg Tyr Arg Gly Arn Ser Tyr His Asp Arg Lys Ser
 195 200 205
 Lys Val Asp Leu Asp Arg Leu Asn Asp Asp Ala Lys Arg Tyr Ser Cys
 210 215 220

FIG. 6A

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Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg Glu Glu Leu Lus Leu Ala
 225 230 235 240
 Asn Val Val Phe Phe Pro Arg Cys Leu Leu Val Gln Arg Cys Asn Ser
 245 250 255
 Asn Cys Gly Cys Gly Thr Val Asn Trp Arg Ser Cys Thr Cys Asn Ser
 260 265 270
 Gly Lys Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro Gly
 275 280 285
 His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile
 290 295 300
 Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 305 310 315 320
 Pro Arg

FIG. 6B

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cgctcggaaa gttcagcatg caggaagltt ggggagagct cggcgattag cacagcgacc 60
 cgggccagcg cagggcgagc gcagggcgcg agagcgagg gcggcgggc gtcggtcccg 120
 ggagcagaac ccggcttttt ctggagcga cgctgtctct agtcgtgat cccaa 175
 atg cac cgg ctc atc ttt glc tac act cta atc tgc gca aac ttt tgc 223
 Met His Arg Leu Ile Phe Val Tyr Thr Leu Ile Cys Ala Asn Phe Cys
 1 5 10 15
 agc tgt cgg gac act tct gca acc ccg cag agc gca tcc atc aaa gct 271
 Ser Cys Arg Asp The Ser Ala Thr Pro Gln Ser Ala Ser Ile Lus Ala
 20 25 30
 ttg cgc aac gcc aac ctc agg cga gat gag agc aat cac ctc aca gac 319
 Leu Arg Asn Ala Asn Leu Arg Arg Asp Glu Ser Asn His Leu Thr Asp
 35 40 45
 ttg tac cga aga gat gag acc atc cag gtg aaa gga aac ggc tac gtg 367
 Leu Tyr Arg Arg Asp Glu Thr Ile Gln Val Lys Gly Asn Gly Tyr Val
 50 55 60
 cag agt cct aga ttc ccg aac agc tac ccc agg aac ctg ctc ctg aca 415
 Gln Ser Pro Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Thr
 65 70 75 80
 tgg cgg ctt cac tct cag gag aat aca cgg ata cat cta gtg ttt gac 463
 Trp Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp
 85 90 95
 aat cag ttt gga tta gag gaa gca gaa aat gat atc tgt agg tat gat 511
 Asn Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp
 100 105 110
 ttt gtg gaa gtt gaa gat ata tcc gaa acc agt acc att att aga gga 559
 Phe Val Glu Val Glu Asp Ile Ser Glu Thr Ser The Ile Ile Arg Gly
 115 120 125
 cga tgg tgt gga cac aag gaa gtt cct cca agg ata aaa tca aga acg 607
 Arg Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr
 130 135 140
 aac caa att aaa atc aca ttc aag tcc gat gac tac ttt gtg gct aaa 655
 Asn Gln Ile Lys Ile Thr Phe Pys Ser Asp Asp Tyr Phe Val Ala Lys
 145 150 155 160

FIG. 7A

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cct gga ttc aag att tat tat tct ttg ctg gaa gat ttc caa ccc gca Fro Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro Ala	703
165 170 175	
gca gct tta gag acc aac tgg gaa tct glc aca agc tct att tca ggg Ala Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser Gly	751
180 185 190	
gta tcc tat aac tct cca tca gta act gat ccc act ctg att gag gat Val Ser Tyr Asn Ser Pro Ser Val Thr Asp Pro Thr Leu Ile Ala Asp	799
195 200 205	
gct ctg gac aaa aaa att gca gaa ttt gat aca gtg gaa gat ctg ctc Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp Thr Val Glu Asp Leu Lei	847
210 215 220	
aag tac ttc aat cca gag tca tgg caa gaa gat ctt gag aat atg tat Lys thr Phe Asn Pro Glu Ser Trp Gln Glu Asp Leu Flu Asn Met Tyr	895
225 230 235 240	
ctg gac acc cct cgg tat cga ggc agg tca tac cat gac cgg aag tca Leu Asp Thr Pro Arg Tyr Arg Gly Arg Ser Tyr His Asp Arg Lys Ser	943
245 250 255	
aaa gtt gac ctg gat agg ctc aat gat gat gcc aag cgt tac agt tgc Lys Val Asl Leu Asp Arg Leu Asn Asp Asp Ala Lus Arg Tyr Ser Cys	991
260 265 270	
act ccc agg aat tac tcg gtc aat ata aga gaa gag ctg aag ttg gcc Thr Pro Arg Asn Try Ser Val Asn IIs Arg Glu Glu Leu Lys Leu Ala	1039
275 280 285	
aat gtg gtc ttc ttt cca cgt tgc ctc ctc gtg cag cgc tgt gga gga Asn Val Val Phe Phe Pro Arg Cys Leu Leu Val Gln Arg Cys Gly Gly	1087
290 295 300	
aat tgt ggc tgt gga act gtc aac tgg agg tcc tgc aca tgc aat tca Asn Cys Gly Cys Gly Thr Val Asn Trg Arg Ser Cys Thr Cys Asn Ser	1135
305 310 315 320	
ggg aaa acc gtg aaa aag tat cat gag gta tta cag ttt gag cct ggc Gly Lys Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro Gly	1183
325 330 335	

FIG. 7B

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cac atc aag agg agg ggt aga gct aag acc atg gct cta gtt gac atc 1231
 His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile
 340 345 350

cag ttg gat cac cat gaa cga tgc gat tgt atc tgc agc tca aga cca 1279
 Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 355 360 365

cct cga taagagaatg tgcacatcct tacatttaagc ctgaagaac ctttagttta 1335
 Pro Arg
 370

aggggggtga gataagagac ccttttccca ccagcaacca aacttactac tagcctgcaa 1396
 tgcattgaac acaagtgggt gctgagcttc agccttgctt tgltaatgcc atggcaagla 1455
 gaaaggata tcatcaactt ctatacctaa gaatatagga ttgcatttaa taataglggt 1515
 tgagggtata tatgcacaaa cacacacaga aatatattca tglctatgtg tatatagatc 1575
 aatgtttttt ttgggtatat ataaccaggt acaccagagc ttacatatgt ttgagltaga 1635
 ctcttaaaat cctttgccaa aataagggat ggtcaaatat atgaacatg tctttagaaa 1695
 atttaggaga taaatttatt tttaaatitt gaacacacaa acaattitga atcttgctct 1755
 cttaagagaa gcatcttcta tottaaaaat caaagatga ggccttctta catatacatc 1815
 ttagltgatt attaaaaaag gaaaaagggt tccagagaaa aggccaatcc ctaagcattt 1875
 ttccatgag aagcactgca tacttacctc tglggactgt aataacctgt ctccaaaacc 1935
 atgccataat aatataagtg ctttagaaat taatcatlg tgttttttat gcatlttgct 1995
 gaggcattct tattcatlta acacctatct caaaaactta cttagaagggt tttttattat 2055
 agtcttaca aagacaatgt ataagctgta acagaatttt gaattgtttt tctttgcaa 2115
 accctccac aaaagcaaat cctttcaaga atggcatggg cattctgtat gaacctttcc 2175
 agatgggtgt cagtgaaga tgtgggtagt tgagaactta aaaagtgaac attgaacat 2235
 cgacgtaact ggaaaccg????????2253

FIG. 7C

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Met His Arg Leu Ile Phe Val Tyr Thr Leu Ile Cys Ala Asn Phe
 1 5 10 15
 Cys Ser Cys Arg Asp Thr Ser Ala Thr Pro Gln Ser Ala Ser Ile Lys
 20 25 30
 Ala Leu Arg Asn Ala Asn Leu Arf Arg Asp Glu Ser Asn His Leu Thr
 35 40 45
 Asp Leu Tyr Arg Arg Asp Glu Thr Ile Gln Val Lys Gly Asn Gly Tyr
 50 55 60
 Val Gln Ser Pro Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Leu
 65 70 75
 Thr Trp Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe
 80 85 90 95
 Asp Asn Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr
 100 105 110
 Asp Phe Val Glu Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg
 115 120 125
 Gly Arg Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg
 130 135 140
 Thr Asn Gln Ile Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala
 145 150 155
 Lys Pro Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro
 160 165 170 175
 Ala Ala Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser
 180 185 190
 Gly Val Ser Tyr Asn Ser pro Ser Val Thr Asp Pro Thr Leu Ile Ala
 195 200 205
 Asp Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp Thr Val Glu Asp Leu
 210 215 220
 Leu Lys Thr Phe Asn Pro Glu Ser trp Gln Glu Asp Leu Glu Asn Met
 225 230 235

FIG. 8A

SUBSTITUTE SHEET (rule 26)

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Tyr Leu Ash Thr Pro Arg Tyr Arg Gly Arg Ser Tyr His Asp Arg Lys
 240 245 250 255
 Ser Lys Val Asp Leu Asp Arg Leu Asn Asp Asp Ala Lys Arg Tyr Ser
 260 265 270
 Cys Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg Glu Glu Leu Lys Leu
 275 280 285
 Ala Asn Val Val Phe Phe Pro Arg Cys Leu Leu Val Gln Arg Cys Gly
 290 295 300
 Gly Asn Cys Cys Cys Gly Thr Val Asn Trp Arg Ser Cys Thr Cus Asn
 305 310 315
 Ser Gly Lys Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro
 320 325 330 335
 Glu His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp
 340 345 350
 Ila Glm Leu Asp His His Gln Arg Cys Asp Cys Ile Cys Ser Ser Arg
 355 360 365
 Pro Pro Arg
 370

FIG. 8B

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FIG. 9A

FIG. 9B

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VEGF 165	P L M R C C G G	---	C C N D E G L E	C V P T I E E S N I T M Q I M R I I K	---	P R 112
PIGF-2	S L L R C T G	---	C C G D E D L H C	V P V E T A N V T M Q L L K I I A	---	S G 112
VEGF-B167	T V Q R C C G G	---	C C P D D G L E C	V P T G Q H Q V R M Q I L M I R Y	---	P S 108
Pox Orf VEGF	T L M R C C G G	---	C C N D E S L E C	V P T E E V N V S M E L L G A S G S G S N 99		
VEGF-C	S V Y R C C G G	---	C C N S E G L Q C	M N T S T S Y L S K I L F E I T V	---	P L 192
VEGF-D	N V F R C C G G	---	C C N E E G V M C	M N T S T S Y I S K Q L F E I S V	---	P L 177
PCCF-A	E V K R C T G	---	C C N T S S V K C	Q P S R V H H R S V K V A K V E Y V R K K 161		
PCCF-B	E V Q R C S G	---	C C N N R N V Q C	R P T C V Q L R P V Q V R K I I E I V R K K 162		
hPCGF-D	L V Q R C C G G	N C G	C G T V N W R S C	T C N S G K T V K K Y H E V L Q F E P C H 289		
VEGF 165	Q G Q	---	H I G E M S F L Q H N K	C E C R P P K K	---	135
PIGF-2	D R F	---	S Y V E L T P S Q E V R	C E C R P P L R E	---	K M K P E 140
VEGF-B167	S Q L	---	G E M S L E E H S Q	C E C K P K K	---	D S A V 133
Pox Orf VEGF	G M Q	---	R L S F V E H K K	C D C R P R F T	---	T T 121
VEGF-C	S O G	---	P K P V T I S F A N H T S	C R C M S K L D	---	V Y K Q V H 222
VEGF-D	Y S V	---	P E L V P V K I A N H T G	C K C L P T G P	---	R H P Y 205
PCCF-A	P K L	---	K E V Q V R L E E H L E	C A C A T I T S L N P D Y R E E D T 193		
PCCF-B	P I F	---	K K A T V T L E D E L A	C K C E T V A A A R P V T R S P G 194		
hPDGF-D	I K R R G R A K T M A L V D I Q L D H H E R	---	C D C I C S S R P P R			322

FIG. 9C

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VEGF 165	DRA	Q	E	N	P	C	G	P	C	S	S	E	R	R	K	K	L	F	V	Q	D	P	Q	T	C	K	C	S	C	K	N	I	D	S	-	R	C	173		
PIGF-2	R	R	R	P	K	C	R	G	K	R	R	E	N	Q	R	P	T	D	C	H	L	C	G	L	A	V	P	R								170				
VEGF-B167	K	P	D	S	P	R	P	L	C	P	R	C	T	Q	H	H	Q	R	P	D	P	R	T	-	-	-	C	R	C	R	R	R	S	P	L	R	C	169		
Pox Orf VEGF	P	P	T	T	T	R	P	P	R	R																														
VEGF-C	S	I	I	R	S	L	P	A	T	-	L	P	Q	C	Q	A	A	N	R	I	C	P	T	N	Y	M	N	N	H	I	C	R	C	L	A	Q	E	D	261	
VEGF-D	S	I	I	R	S	I	O	I	P	E	E	D	E	C	P	H	S	K	K	L	C	P	I	D	M	L	W	D	N	T	K	C	K	C	V	L	O	D	E	245
PCGF-A	G	R	P	R	E	S	G	K	K	R	K	R	L	K	P	T																								
PCGF-B	G	S	Q	E	Q	R	A	K	T	P	Q	T	R	V	T	I	R	T	V	R	R	P	P	K	G	K	H	R	K	F	K	H	T	H	D	K	T	A	234	
hPDGF-D																																				322				
VEGF 165	K	A	R	Q	L	E	L	N	E	R	T	C	R	C	D	K	P	R																			152			
PIGF-2																																			170					
VEGF-B167	Q	G	R	G	L	E	L	N	P	D	T	C	R	C	R	K	L	R																		188				
Pox Orf VEGF																																			133					
VEGF-C	F	M	F	S	S	D	A	G	D	S	I	D	G	F	E	D	I	C	G	P	N	K	E	L	D	E	E	T	C	Q	C	V	C	R	A	G	L	R	P	301
VEGF-D	-	T	P	L	F	G	T	E	D	H	S	Y	L	O	E	P	T	L	C	G	P	H	M	T	F	D	E	D	R	-	-	-	-	-	-	-	-	274		
PDGF-A																																								
PDGF-B	L	K	E	T	L	G	A																													211				
hPDGF-D																																				241				
																																			322					

FIG. 9D

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VEGF 165	192
PIGF-2	170
VEGF-B167	188
Pox Orf VEGF	133
VEGF-C	341
VEGF-D	295
PCCF-A	211
PCCF-B	241
hPDGF-D	322
VEGF 165	192
PIGF-2	170
VEGF-B167	188
Pox Orf VEGF	133
VEGF-C	381
VEGF-D	317
PCCF-A	211
PCCF-B	241
hPDGF-D	322

A S C G P H K E L D R N S C Q C V C K N K L F P S Q C G A N R E F D E N T C Q C
 - - - - - C E C V C K A P C P G D L I O H P E N - - - - - C S C

V C K R T C P R N O P L N P G K C A C E C T E S P Q K C L L K G K K F H H Q T C
 F E C K E S L E S C C O K H K I - - - - - F H P D T C

FIG. 9E

VEGF 165
 PlGF-2
 VEGF-B167
 Pox Orf VEGF
 VEGF-C
 VEGF-D
 PCGF-A
 PCGF-B
 hPDGF-D

VEGF 165
 PlGF-2
 VEGF-B167
 Pox Orf VEGF
 VEGF-C
 VEGF-D
 PCGF-A
 PCGF-B
 hPDGF-D

SCYR
 SCEDR
 P
 C
 T
 N
 R
 Q
 K
 A
 C
 E
 P
 G
 F
 S
 Y
 S
 E
 E
 V
 C
 R
 C
 V
 P
 S
 Y
 W
 K
 R
 P
 Q
 M
 S
 C
 P
 F
 H
 T
 R
 T
 C
 A
 S
 R
 K
 P
 F
 A
 C
 G
 K
 H
 W
 R
 P
 K
 E
 T
 R
 A
 O
 G
 L
 Y
 S
 O
 E

VEGF 165
 PlGF-2
 VEGF-B167
 Pox Orf VEGF
 VEGF-C
 VEGF-D
 PCGF-A
 PCGF-B
 hPDGF-D

VEGF 165
 PlGF-2
 VEGF-B167
 Pox Orf VEGF
 VEGF-C
 VEGF-D
 PCGF-A
 PCGF-B
 hPDGF-D

NP

FIG. 9F

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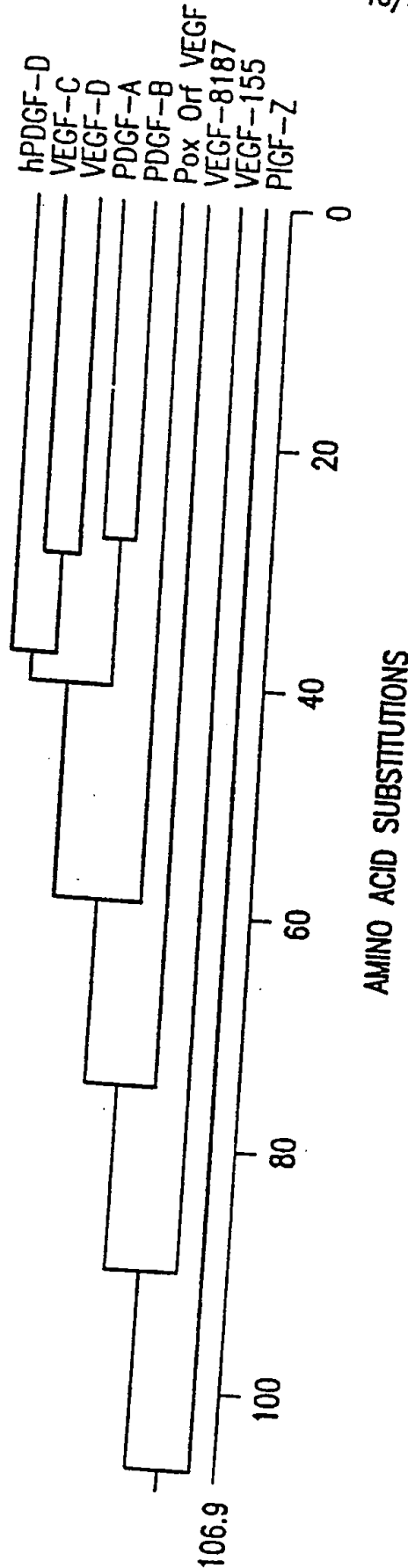


FIG. 10

hPCDF-D CUB
hBMP-1 CUB1
hBMP-1 CUB2
hBMP-1 CUB3
Neuropilin CUB1
Neuropilin CUB2
hPCDF-D CUB
hBMP-1 CUB1
hBMP-1 CUB2
hBMP-1 CUB3
Neuropilin CUB1
Neuropilin CUB2
hPCDF-D CUB
hBMP-1 CUB1
hBMP-1 CUB2
hBMP-1 CUB3
Neuropilin CUB1
Neuropilin CUB2
hPCDF-D CUB
hBMP-1 CUB1
hBMP-1 CUB2
hBMP-1 CUB3
Neuropilin CUB1
Neuropilin CUB2

FIG. 11

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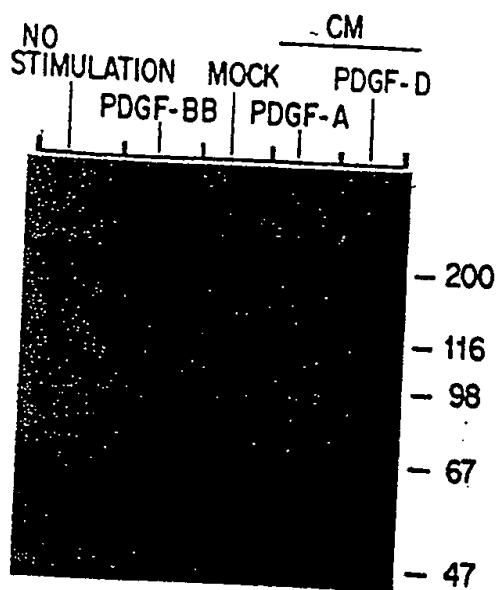


FIG. 12

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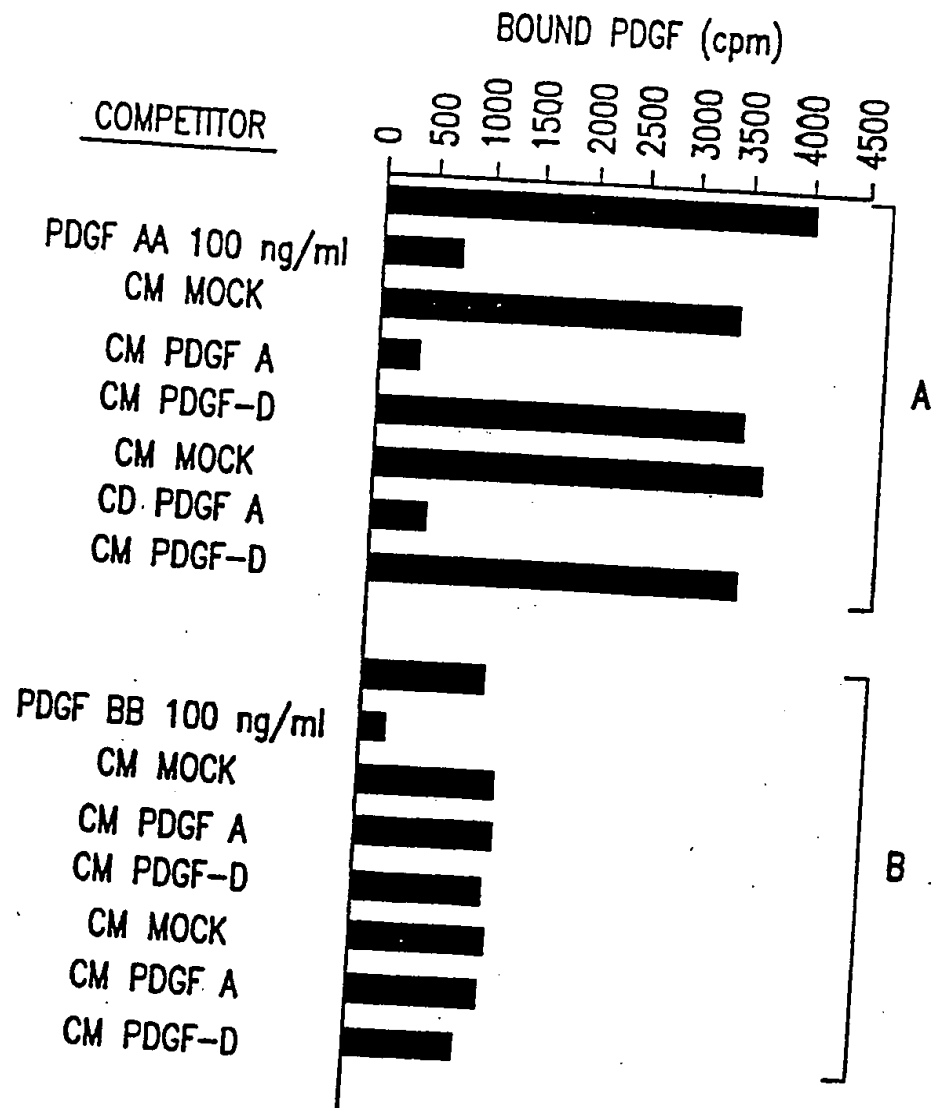


FIG. 13

SEQUENCE LISTING

<110> ERIKSSON, Ulf
AASE, Karin
LEE, Xuri
PONTUN, Annica
UUTELA, Marko
ALITALO, Kari
OESTMAN, Arne
HELDIN, Carl-Henrik

<120> PLATELET-DERIVED GROWTH FACTOR D, DNA CODING
THEREFOR, AND USES THEREOF

<130> Ulf Eriksson et al 1064-44833PC

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<150> 60/107,852

<151> 1998-11-10

<150> 60/113,997

<151> 1999-12-28

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<151> 1999-10-05

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<170> PatentIn Ver. 2.0

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aagaccatgg ctctagtga catccagttg gatcaccatg aacgatgtga ttgtatctgc 180
 agctcaagac cacctcgata agagaatgtg cacatcctta cattaagcct gaaagaacca 240
 ttagtttaag gaggggtgaga taagagaccc ttttcctacc agcaaccaga cttactacta 300
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 20 25 30
 His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile
 35 40 45
 Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 50 55 60
 Pro Arg
 65

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 gacaaaaaaaa ttgcagaatt tgatacagtg gaagatctgc tcaagtactt caatccagag 180
 tcatggcaag aagatcttga gaatatgtat ctggacaccc ctcggtatcg aggcagggtca 240
 taccatgacc ggaagtcaaa agttgacctg gatagggtca atgatgatgc caagcggttac 300
 agttgcactc ccaggaatta ctcggtcaat ataagagaag agctgaagtt ggccaatgtg 360

gtcttctttc cacgttgcoct cctcgtgcag cgctgtggag gaaattgtgg ctgtggaact 420
 gtcaaactgg agtcctgcac atgcaattca gggaaaaccg tgaaaaagta tcatgaggta 480
 ttacagtttg agcctggcca catcaagagg aggggtagag ctaagaccat ggctctagtt 540
 gacatccagt tggatcacca tgaacgatgc gattgtatct gcagctcaag accacctcga 600
 taagagaatg tgcacatcct tacattaagc ctgaaagaac ctttagttta aggagggtga 660
 gataagagac ccttttcccta ccagcaaccc 690

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<213> Homo sapiens

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 20 25 30

Pro Thr Leu Ile Ala Asp Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp
 35 40 45

Thr Val Glu Asp Leu Leu Lys Tyr Phe Asn Pro Glu Ser Trp Gln Glu
 50 55 60

Asp Leu Glu Asn Met Tyr Leu Asp Thr Pro Arg Tyr Arg Gly Arg Ser
 65 70 75 80

Tyr His Asp Arg Lys Ser Lys Val Asp Leu Asp Arg Leu Asn Asp Asp
 85 90 95

Ala Lys Arg Tyr Ser Cys Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg
 100 105 110

Glu Glu Leu Lys Leu Ala Asn Val Val Phe Phe Pro Arg Cys Leu Leu
 115 120 125

Val Gln Arg Cys Gly Gly Asn Cys Gly Cys Gly Thr Val Lys Leu Glu
 130 135 140

Ser Cys Thr Cys Asn Ser Gly Lys Thr Val Lys Lys Tyr His Glu Val

145
 150
 155
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 Leu Gln Phe Glu Pro Gly His Ile Lys Arg Arg Gly Arg Ala Lys Thr
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 Ile Cys Ser Ser Arg Pro Pro Arg
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 cag agt cct aga ttc ccg aac agc tac ccc agg aac ctg ctc ctg aca 96
 Gln Ser Pro Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Thr
 20 25 30
 tgg cgg ctt cac tct cag gag aat aca cgg ata cag cta gtg ttt gac 144
 Trp Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp
 35 40 45
 aat cag ttt gga tta gag gaa gca gaa aat gat atc tgt agg tat gat 192
 Asn Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp
 50 55 60
 ttt gtg gaa gtt gaa gat ata tcc gaa acc agt acc att att aga gga 240
 Phe Val Glu Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg Gly
 65 70 75 80
 cga tgg tgt gga cac aag gaa gtt cct cca agg ata aaa tca aga acg 288
 Arg Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr
 85 90 95
 aac caa att aaa atc aca ttc aag tcc gat gac tac ttt gtg gct aaa 336
 Asn Gln Ile Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala Lys

100	105	110	
cct gga ttc aag att tat tat tct ttg ctg gaa gat ttc caa ccc gca			384
Pro Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro Ala			
115	120	125	
gca gct tca gag acc aac tgg gaa tct gtc aca agc tct att tca ggg			432
Ala Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser Gly			
130	135	140	
gta tcc tat aac tct cca tca gta acg gat ccc act ctg att gcg gat			480
Val Ser Tyr Asn Ser Pro Ser Val Thr Asp Pro Thr Leu Ile Ala Asp			
145	150	155	160
gct ctg gac aaa aaa att gca gaa ttt gat aca gtg gaa gat ctg ctc			528
Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp Thr Val Glu Asp Leu Leu			
165	170	175	
aag tac ttc aat cca gag tca tgg caa gaa gat ctt gag aat atg tat			576
Lys Tyr Phe Asn Pro Glu Ser Trp Gln Glu Asp Leu Glu Asn Met Tyr			
180	185	190	
ctg gac acc cct cgg tat cga ggc agg tca tac cat gac cgg aag tca			624
Leu Asp Thr Pro Arg Tyr Arg Gly Arg Ser Tyr His Asp Arg Lys Ser			
195	200	205	
aaa gtt gac ctg gat agg ctc aat gat gat gcc aag cgt tac agt tgc			672
Lys Val Asp Leu Asp Arg Leu Asn Asp Asp Ala Lys Arg Tyr Ser Cys			
210	215	220	
act ccc agg aat tac tcg gtc aat ata aga gaa gag ctg aag ttg gcc			720
Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg Glu Glu Leu Lys Leu Ala			
225	230	235	240
aat gtg gtc ttc ttt cca cgt tgc ctc ctc gtg cag cgc tgt gga gga			768
Asn Val Val Phe Phe Pro Arg Cys Leu Leu Val Gln Arg Cys Gly Gly			
245	250	255	
aat tgt ggc tgt gga act gtc aac tgg agg tcc tgc aca tgc aat tca			816
Asn Cys Gly Cys Gly Thr Val Asn Trp Arg Ser Cys Thr Cys Asn Ser			
260	265	270	
ggg aaa acc gtg aaa aag tat cat gag gta tta cag ttt gag cct ggc			864
Gly Lys Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro Gly			
275	280	285	
cac atc aag agg agg ggt aga gct aag acc atg gct cta gtt gac atc			912
His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile			

290

295

300

cag ttg gat cac cat gaa cga tgc gat tgt atc tgc agc tca aga cca 960
 Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 305 310 315 320

cct cga taagagaatg tgcacatcct tacattaagc ctgaaagaac ctttagttta 1016
 Pro Arg

aggaggggtga gataagagac ccttttccta ccagcaacca aacttactac tagcctgcaa 1076
 tgcaatgaac acaagtgggt gctgagtctc agccttgctt tgtaaatgcc atggcaagta 1136
 gaaaggtata tcatcaactt ctatacctaa gaatatagga ttgcatttaa taatagtgtt 1196
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 cttaaagaaa gcatcttgta tattaaaaat caaaagatga ggctttctta catatacatc 1496
 ttagttgatt attaaaaaag gaaaaagggt tccagagaaa aggccaatac ctaagcattt 1556
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 gaggcacctt tattcattta acacctatct caaaaactta cttagaagggt tttttattat 1736
 agtcctacaa aagacaatgt ataagctgta acagaatttt gaattgtttt tctttgcaaa 1796
 acccctccac aaaagcaaat cctttcaaga atggcatggg cattctgtat gaacctttcc 1856
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 cgacgtaact ggaaaccg 1934

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<211> 322

<212> PRT

<213> Homo sapiens

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 20 25 30
 Trp Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp
 35 40 45
 Asn Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp
 50 55 60
 Phe Val Glu Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg Gly
 65 70 75 80
 Arg Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr
 85 90 95
 Asn Gln Ile Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala Lys
 100 105 110
 Pro Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro Ala
 115 120 125
 Ala Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser Gly
 130 135 140
 Val Ser Tyr Asn Ser Pro Ser Val Thr Asp Pro Thr Leu Ile Ala Asp
 145 150 155 160
 Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp Thr Val Glu Asp Leu Leu
 165 170 175
 Lys Tyr Phe Asn Pro Glu Ser Trp Gln Glu Asp Leu Glu Asn Met Tyr
 180 185 190
 Leu Asp Thr Pro Arg Tyr Arg Gly Arg Ser Tyr His Asp Arg Lys Ser
 195 200 205
 Lys Val Asp Leu Asp Arg Leu Asn Asp Asp Ala Lys Arg Tyr Ser Cys
 210 215 220
 Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg Glu Glu Leu Lys Leu Ala
 225 230 235 240
 Asn Val Val Phe Phe Pro Arg Cys Leu Leu Val Gln Arg Cys Gly Gly
 245 250 255

Asn Cys Gly Cys Gly Thr Val Asn Trp Arg Ser Cys Thr Cys Asn Ser
 260 265 270

Gly Lys Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro Gly
 275 280 285

His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile
 290 295 300

Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 305 310 315 320

Pro Arg

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<212> DNA

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cgggccagcg cagggcgagc gcaggcggcg agagcgcagg gcggcgcgcc gtcgggtccc 120

ggagcagaac ccggcttttt cttggagcga cgctgtctct agtcgtgat ccaa atg 178
 Met

1

cac cgg ctc atc ttt gtc tac act cta atc tgc gca aac ttt tgc agc 226
 His Arg Leu Ile Phe Val Tyr Thr Leu Ile Cys Ala Asn Phe Cys Ser
 5 10 15

tgt cgg gac act tct gca acc ccg cag agc gca tcc atc aaa gct ttg 274
 Cys Arg Asp Thr Ser Ala Thr Pro Gln Ser Ala Ser Ile Lys Ala Leu
 20 25 30

cgc aac gcc aac ctc agg cga gat gag agc aat cac ctc aca gac ttg 322
 Arg Asn Ala Asn Leu Arg Arg Asp Glu Ser Asn His Leu Thr Asp Leu
 35 40 45

tac cga aga gat gag acc atc cag gtg aaa gga aac ggc tac gtg cag 370
 Tyr Arg Arg Asp Glu Thr Ile Gln Val Lys Gly Asn Gly Tyr Val Gln

50	55	60	65	
agt cct aga ttc ccg aac agc tac ccc agg aac ctg ctc ctg aca tgg				418
Ser Pro Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Leu Thr Trp				
70	75	80		
cgg ctt cac tct cag gag aat aca cgg ata cag cta gtg ttt gac aat				466
Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp Asn				
85	90	95		
cag ttt gga tta gag gaa gca gaa aat gat atc tgt agg tat gat ttt				514
Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp Phe				
100	105	110		
gtg gaa gtt gaa gat ata tcc gaa acc agt acc att att aga gga cga				562
Val Glu Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg Gly Arg				
115	120	125		
tgg tgt gga cac aag gaa gtt cct cca agg ata aaa tca aga acg aac				610
Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr Asn				
130	135	140	145	
caa att aaa atc aca ttc aag tcc gat gac tac ttt gtg gct aaa cct				658
Gln Ile Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala Lys Pro				
150	155	160		
gga ttc aag att tat tat tct ttg ctg gaa gat ttc caa ccc gca gca				706
Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro Ala Ala				
165	170	175		
gct tca gag acc aac tgg gaa tct gtc aca agc tct att tca ggg gta				754
Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser Gly Val				
180	185	190		
tcc tat aac tct cca tca gta acg gat ccc act ctg att gcg gat gct				802
Ser Tyr Asn Ser Pro Ser Val Thr Asp Pro Thr Leu Ile Ala Asp Ala				
195	200	205		
ctg gac aaa aaa att gca gaa ttt gat aca gtg gaa gat ctg ctc aag				850
Leu Asp Lys Lys Ile Ala Glu Phe Asp Thr Val Glu Asp Leu Leu Lys				
210	215	220	225	
tac ttc aat cca gag tca tgg caa gaa gat ctt gag aat atg tat ctg				898
Tyr Phe Asn Pro Glu Ser Trp Gln Glu Asp Leu Glu Asn Met Tyr Leu				
230	235	240		
gac acc cct cgg tat cga ggc agg tca tac cat gac cgg aag tca aaa				946
Asp Thr Pro Arg Tyr Arg Gly Arg Ser Tyr His Asp Arg Lys Ser Lys				

10

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<213> Homo sapiens

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 20 25 30

Leu Arg Asn Ala Asn Leu Arg Arg Asp Glu Ser Asn His Leu Thr Asp
 35 40 45

Leu Tyr Arg Arg Asp Glu Thr Ile Gln Val Lys Gly Asn Gly Tyr Val
 50 55 60

Gln Ser Pro Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Leu Thr
 65 70 75 80

Trp Arg Leu His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp
 85 90 95

Asn Gln Phe Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp
 100 105 110

Phe Val Glu Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg Gly
 115 120 125

Arg Trp Cys Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr
 130 135 140
 Asn Gln Ile Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala Lys
 145 150 155 160
 Pro Gly Phe Lys Ile Tyr Tyr Ser Leu Leu Glu Asp Phe Gln Pro Ala
 165 170 175
 Ala Ala Ser Glu Thr Asn Trp Glu Ser Val Thr Ser Ser Ile Ser Gly
 180 185 190
 Val Ser Tyr Asn Ser Pro Ser Val Thr Asp Pro Thr Leu Ile Ala Asp
 195 200 205
 Ala Leu Asp Lys Lys Ile Ala Glu Phe Asp Thr Val Glu Asp Leu Leu
 210 215 220
 Lys Tyr Phe Asn Pro Glu Ser Trp Gln Glu Asp Leu Glu Asn Met Tyr
 225 230 235 240
 Leu Asp Thr Pro Arg Tyr Arg Gly Arg Ser Tyr His Asp Arg Lys Ser
 245 250 255
 Lys Val Asp Leu Asp Arg Leu Asn Asp Asp Ala Lys Arg Tyr Ser Cys
 260 265 270
 Thr Pro Arg Asn Tyr Ser Val Asn Ile Arg Glu Glu Leu Lys Leu Ala
 275 280 285
 Asn Val Val Phe Phe Pro Arg Cys Leu Leu Val Gln Arg Cys Gly Gly
 290 295 300
 Asn Cys Gly Cys Gly Thr Val Asn Trp Arg Ser Cys Thr Cys Asn Ser
 305 310 315 320
 Gly Lys Thr Val Lys Lys Tyr His Glu Val Leu Gln Phe Glu Pro Gly
 325 330 335
 His Ile Lys Arg Arg Gly Arg Ala Lys Thr Met Ala Leu Val Asp Ile
 340 345 350
 Gln Leu Asp His His Glu Arg Cys Asp Cys Ile Cys Ser Ser Arg Pro
 355 360 365
 Pro Arg
 370

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 Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln
 35 40 45
 Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu
 50 55 60
 Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu
 65 70 75 80
 Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro
 85 90 95
 Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His
 100 105 110
 Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys
 115 120 125
 Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Asn Pro Cys Gly
 130 135 140
 Pro Cys Ser Ser Glu Arg Arg Lys His Leu Phe Val Gln Asp Pro Gln
 145 150 155 160

Thr Cys Lys Cys Ser Cys Lys Asn Thr Asp Ser Arg Cys Lys Ala Arg
 165 170 175

Gln Leu Glu Leu Asn Glu Arg Thr Cys Arg Cys Asp Lys Pro Arg Arg
 180 185 190

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 20 25 30

Asn Gly Ser Ser Glu Val Glu Val Val Pro Phe Gln Glu Val Trp Gly
 35 40 45

Arg Ser Tyr Cys Arg Ala Leu Glu Arg Leu Val Asp Val Val Ser Glu
 50 55 60

Tyr Pro Ser Glu Val Glu His Met Phe Ser Pro Ser Cys Val Ser Leu
 65 70 75 80

Leu Arg Cys Thr Gly Cys Cys Gly Asp Glu Asp Leu His Cys Val Pro
 85 90 95

Val Glu Thr Ala Asn Val Thr Met Gln Leu Leu Lys Ile Arg Ser Gly
 100 105 110

Asp Arg Pro Ser Tyr Val Glu Leu Thr Phe Ser Gln His Val Arg Cys
 115 120 125

Glu Cys Arg Pro Leu Arg Glu Lys Met Lys Pro Glu Arg Arg Arg Pro
 130 135 140

Lys Gly Arg Gly Lys Arg Arg Arg Glu Asn Gln Arg Pro Thr Asp Cys
 145 150 155 160

His Leu Cys Gly Asp Ala Val Pro Arg Arg
 165 170

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 <212> PRT
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<400> 12
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 Ala Pro Ala Gln Ala Pro Val Ser Gln Pro Asp Ala Pro Gly His Gln
 20 25 30
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 35 40 45
 Pro Arg Glu Val Val Val Pro Leu Thr Val Glu Leu Met Gly Thr Val
 50 55 60
 Ala Lys Gln Leu Val Pro Ser Cys Val Thr Val Gln Arg Cys Gly Gly
 65 70 75 80
 Cys Cys Pro Asp Asp Gly Leu Glu Cys Val Pro Thr Gly Gln His Gln
 85 90 95
 Val Arg Met Gln Ile Leu Met Ile Arg Tyr Pro Ser Ser Gln Leu Gly
 100 105 110
 Glu Met Ser Leu Glu Glu His Ser Gln Cys Glu Cys Arg Pro Lys Lys
 115 120 125
 Lys Asp Ser Ala Val Lys Pro Asp Ser Pro Arg Pro Leu Cys Pro Arg
 130 135 140
 Cys Thr Gln His His Gln Arg Pro Asp Pro Arg Thr Cys Arg Cys Arg
 145 150 155 160
 Cys Arg Arg Arg Ser Phe Leu Arg Cys Gln Gly Arg Gly Leu Glu Leu
 165 170 175
 Asn Pro Asp Thr Cys Arg Cys Arg Lys Leu Arg Arg
 180 185

<210> 13
 <211> 364
 <212> PRT

<213> Homo sapiens

<400> 13

Met Glu Thr Leu Tyr Ser Leu Glu Leu Glu Val Ala Leu Gly Leu Tyr
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 Ile Leu Glu Leu Glu Val Ala Leu Ala Leu Ala Val Ala Leu Cys Tyr
 20 25 30
 Ser Leu Glu His Ile Ser Gly Leu Asn Thr Tyr Arg Leu Glu Leu Glu
 35 40 45
 Ala Ser Asn Ala Leu Ala Ala Ser Pro Ser Glu Arg Ala Ser Asn Thr
 50 55 60
 His Arg Leu Tyr Ser Gly Leu Tyr Thr Arg Pro Ser Glu Arg Gly Leu
 65 70 75 80
 Val Ala Leu Leu Glu Leu Tyr Ser Gly Leu Tyr Ser Glu Arg Gly Leu
 85 90 95
 Cys Tyr Ser Leu Tyr Ser Pro Arg Ala Arg Gly Pro Arg Ile Leu Glu
 100 105 110
 Val Ala Leu Val Ala Leu Pro Arg Val Ala Leu Ser Glu Arg Gly Leu
 115 120 125
 Thr His Arg His Ile Ser Pro Arg Gly Leu Leu Glu Thr His Arg Ser
 130 135 140
 Glu Arg Gly Leu Asn Ala Arg Gly Pro His Glu Ala Ser Asn Pro Arg
 145 150 155 160
 Pro Arg Cys Tyr Ser Val Ala Leu Thr His Arg Leu Glu Met Glu Thr
 165 170 175
 Ala Arg Gly Cys Tyr Ser Gly Leu Tyr Gly Leu Tyr Cys Tyr Ser Cys
 180 185 190
 Tyr Ser Ala Ser Asn Ala Ser Pro Gly Leu Ser Glu Arg Leu Glu Gly
 195 200 205
 Leu Cys Tyr Ser Val Ala Leu Pro Arg Thr His Arg Gly Leu Gly Leu
 210 215 220
 Val Ala Leu Ala Ser Asn Val Ala Leu Ser Glu Arg Met Glu Thr Gly
 225 230 235 240

Leu Leu Glu Leu Glu Gly Leu Tyr Ala Leu Ala Ser Glu Arg Gly Leu
 245 250 255

Tyr Ser Glu Arg Gly Leu Tyr Ser Glu Arg Ala Ser Asn Gly Leu Tyr
 260 265 270

Met Glu Thr Gly Leu Asn Ala Arg Gly Leu Glu Ser Glu Arg Pro His
 275 280 285

Glu Val Ala Leu Gly Leu His Ile Ser Leu Tyr Ser Leu Tyr Ser Cys
 290 295 300

Tyr Ser Ala Ser Pro Cys Tyr Ser Ala Arg Gly Pro Arg Ala Arg Gly
 305 310 315 320

Pro His Glu Thr His Arg Thr His Arg Thr His Arg Pro Arg Pro Arg
 325 330 335

Thr His Arg Thr His Arg Thr His Arg Ala Arg Gly Pro Arg Pro Arg
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Ala Arg Gly Ala Arg Gly Ala Arg Gly Ala Arg Gly
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<210> 14

<211> 419

<212> PRT

<213> Homo sapiens

<400> 14

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Ala Leu Leu Pro Gly Pro Arg Glu Ala Pro Ala Ala Ala Ala Phe
 20 25 30

Glu Ser Gly Leu Asp Leu Ser Asp Ala Glu Pro Asp Ala Gly Glu Ala
 35 40 45

Thr Ala Tyr Ala Ser Lys Asp Leu Glu Glu Gln Leu Arg Ser Val Ser
 50 55 60

Ser Val Asp Glu Leu Met Thr Val Leu Tyr Pro Glu Tyr Trp Lys Met
 65 70 75 80

Tyr Lys Cys Gln Leu Arg Lys Gly Gly Trp Gln His Asn Arg Glu Gln
 85 90 95

Ala Asn Leu Asn Ser Arg Thr Glu Glu Thr Ile Lys Phe Ala Ala Ala
 100 105 110
 His Tyr Asn Thr Glu Ile Leu Lys Ser Ile Asp Asn Glu Trp Arg Lys
 115 120 125
 Thr Gln Cys Met Pro Arg Glu Val Cys Ile Asp Val Gly Lys Glu Phe
 130 135 140
 Gly Val Ala Thr Asn Thr Phe Phe Lys Pro Pro Cys Val Ser Val Tyr
 145 150 155 160
 Arg Cys Gly Gly Cys Cys Asn Ser Glu Gly Leu Gln Cys Met Asn Thr
 165 170 175
 Ser Thr Ser Tyr Leu Ser Lys Thr Leu Phe Glu Ile Thr Val Pro Leu
 180 185 190
 Ser Gln Gly Pro Lys Pro Val Thr Ile Ser Phe Ala Asn His Thr Ser
 195 200 205
 Cys Arg Cys Met Ser Lys Leu Asp Val Tyr Arg Gln Val His Ser Ile
 210 215 220
 Ile Arg Arg Ser Leu Pro Ala Thr Leu Pro Gln Cys Gln Ala Ala Asn
 225 230 235 240
 Lys Thr Cys Pro Thr Asn Tyr Met Trp Asn Asn His Ile Cys Arg Cys
 245 250 255
 Leu Ala Gln Glu Asp Phe Met Phe Ser Ser Asp Ala Gly Asp Asp Ser
 260 265 270
 Thr Asp Gly Phe His Asp Ile Cys Gly Pro Asn Lys Glu Leu Asp Glu
 275 280 285
 Glu Thr Cys Gln Cys Val Cys Arg Ala Gly Leu Arg Pro Ala Ser Cys
 290 295 300
 Gly Pro His Lys Glu Leu Asp Arg Asn Ser Cys Gln Cys Val Cys Lys
 305 310 315 320
 Asn Lys Leu Phe Pro Ser Gln Cys Gly Ala Asn Arg Glu Phe Asp Glu
 325 330 335
 Asn Thr Cys Gln Cys Val Cys Lys Arg Thr Cys Pro Arg Asn Gln Pro
 340 345 350

Leu Asn Pro Gly Lys Cys Ala Cys Glu Cys Thr Glu Ser Pro Gln Lys
 355 360 365

Cys Leu Leu Lys Gly Lys Lys Phe His His Gln Thr Cys Ser Cys Tyr
 370 375 380

Arg Arg Pro Cys Thr Asn Arg Gln Lys Ala Cys Glu Pro Gly Phe Ser
 385 390 395 400

Tyr Ser Glu Glu Val Cys Arg Cys Val Pro Ser Tyr Trp Lys Arg Pro
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Gln Met Ser

<210> 15

<211> 358

<212> PRT

<213> Homo sapiens

<400> 15

Met Tyr Gly Glu Trp Gly Met Gly Asn Ile Leu Met Met Phe His Val
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Tyr Leu Val Gln Gly Phe Arg Ser Glu His Gly Pro Val Lys Asp Phe
 20 25 30

Ser Phe Glu Arg Ser Ser Arg Ser Met Leu Glu Arg Ser Glu Gln Gln
 35 40 45

Ile Arg Ala Ala Ser Ser Leu Glu Glu Leu Leu Gln Ile Ala His Ser
 50 55 60

Glu Asp Trp Lys Leu Trp Arg Cys Arg Leu Lys Leu Lys Ser Leu Ala
 65 70 75 80

Ser Met Asp Ser Arg Ser Ala Ser His Arg Ser Thr Arg Phe Ala Ala
 85 90 95

Thr Phe Tyr Asp Thr Glu Thr Leu Lys Val Ile Asp Glu Glu Trp Gln
 100 105 110

Arg Thr Gln Cys Ser Pro Arg Glu Thr Cys Val Glu Val Ala Ser Glu
 115 120 125

Leu Gly Lys Thr Thr Asn Thr Phe Phe Lys Pro Pro Cys Val Asn Val

130
 Phe Arg Cys Gly Gly Cys Cys Asn Glu Glu Gly Val Met Cys Met Asn
 145 150 155 160
 Thr Ser Thr Ser Tyr Ile Ser Lys Gln Leu Phe Glu Ile Ser Val Pro
 165 170 175
 Leu Thr Ser Val Pro Glu Leu Val Pro Val Lys Ile Ala Asn His Thr
 180 185 190
 Gly Cys Lys Cys Leu Pro Thr Gly Pro Arg His Pro Tyr Ser Ile Ile
 195 200 205
 Arg Arg Ser Ile Gln Thr Pro Glu Glu Asp Glu Cys Pro His Ser Lys
 210 215 220
 Lys Leu Cys Pro Ile Asp Met Leu Trp Asp Asn Thr Lys Cys Lys Cys
 225 230 235 240
 Val Leu Gln Asp Glu Thr Pro Leu Pro Gly Thr Glu Asp His Ser Tyr
 245 250 255
 Leu Gln Glu Pro Thr Leu Cys Gly Pro His Met Thr Phe Asp Glu Asp
 260 265 270
 Arg Cys Glu Cys Val Cys Lys Ala Pro Cys Pro Gly Asp Leu Ile Gln
 275 280 285
 His Pro Glu Asn Cys Ser Cys Phe Glu Cys Lys Glu Ser Leu Glu Ser
 290 295 300
 Cys Cys Gln Lys His Lys Ile Phe His Pro Asp Thr Cys Ser Cys Glu
 305 310 315 320
 Asp Arg Cys Pro Phe His Thr Arg Thr Cys Ala Ser Arg Lys Pro Ala
 325 330 335
 Cys Gly Lys His Trp Arg Phe Pro Lys Glu Thr Arg Ala Gln Gly Leu
 340 345 350
 Tyr Ser Gln Glu Asn Pro
 355

<210> 16
 <211> 211
 <212> PRT

<213> Homo sapiens

<400> 16

Met Arg Thr Leu Ala Cys Leu Leu Leu Leu Gly Cys Gly Tyr Leu Ala
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 His Val Leu Ala Glu Glu Ala Glu Ile Pro Arg Glu Val Ile Glu Arg
 20 25 30
 Leu Ala Arg Ser Gln Ile His Ser Ile Arg Asp Leu Gln Arg Leu Leu
 35 40 45
 Glu Ile Asp Ser Val Gly Ser Glu Asp Ser Leu Asp Thr Ser Leu Arg
 50 55 60
 Ala His Gly Val His Ala Thr Lys His Val Pro Glu Lys Arg Pro Leu
 65 70 75 80
 Pro Ile Arg Arg Lys Arg Ser Ile Glu Glu Ala Val Pro Ala Val Cys
 85 90 95
 Lys Thr Arg Thr Val Ile Tyr Glu Ile Pro Arg Ser Gln Val Asp Pro
 100 105 110
 Thr Ser Ala Asn Phe Leu Ile Trp Pro Pro Cys Val Glu Val Lys Arg
 115 120 125
 Cys Thr Gly Cys Cys Asn Thr Ser Ser Val Lys Cys Gln Pro Ser Arg
 130 135 140
 Val His His Arg Ser Val Lys Val Ala Lys Val Glu Tyr Val Arg Lys
 145 150 155 160
 Lys Pro Lys Leu Lys Glu Val Gln Val Arg Leu Glu Glu His Leu Glu
 165 170 175
 Cys Ala Cys Ala Thr Thr Ser Leu Asn Pro Asp Tyr Arg Glu Glu Asp
 180 185 190
 Thr Gly Arg Pro Arg Glu Ser Gly Lys Lys Arg Lys Arg Lys Arg Leu
 195 200 205
 Lys Pro Thr
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<210> 17

<211> 241

<212> PRT

<213> Homo sapiens

<400> 17

Met Asn Arg Cys Trp Ala Leu Phe Leu Ser Leu Cys Cys Tyr Leu Arg
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Leu Val Ser Ala Glu Gly Asp Pro Ile Pro Glu Glu Leu Tyr Glu Met
 20 25 30

Leu Ser Asp His Ser Ile Arg Ser Phe Asp Asp Leu Gln Arg Leu Leu
 35 40 45

His Gly Asp Pro Gly Glu Glu Asp Gly Ala Glu Leu Asp Leu Asn Met
 50 55 60

Thr Arg Ser His Ser Gly Gly Glu Leu Glu Ser Leu Ala Arg Gly Arg
 65 70 75 80

Arg Ser Leu Gly Ser Leu Thr Ile Ala Glu Pro Ala Met Ile Ala Glu
 85 90 95

Cys Lys Thr Arg Thr Glu Val Phe Glu Ile Ser Arg Arg Leu Ile Asp
 100 105 110

Arg Thr Asn Ala Asn Phe Leu Val Trp Pro Pro Cys Val Glu Val Gln
 115 120 125

Arg Cys Ser Gly Cys Cys Asn Asn Arg Asn Val Gln Cys Arg Pro Thr
 130 135 140

Gln Val Gln Leu Arg Pro Val Gln Val Arg Lys Ile Glu Ile Val Arg
 145 150 155 160

Lys Lys Pro Ile Phe Lys Lys Ala Thr Val Thr Leu Glu Asp His Leu
 165 170 175

Ala Cys Lys Cys Glu Thr Val Ala Ala Ala Arg Pro Val Thr Arg Ser
 180 185 190

Pro Gly Gly Ser Gln Glu Gln Arg Ala Lys Thr Pro Gln Thr Arg Val
 195 200 205

Thr Ile Arg Thr Val Arg Val Arg Arg Pro Pro Lys Gly Lys His Arg
 210 215 220

Lys Phe Lys His Thr His Asp Lys Thr Ala Leu Lys Glu Thr Leu Gly
 225 230 235 240

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<210> 18

<211> 121

<212> PRT

<213> Homo sapiens

<400> 18

Ser Tyr His Asp Arg Lys Ser Lys Val Asp Leu Asp Arg Leu Asn Asp
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Asp Ala Lys Arg Tyr Ser Cys Thr Pro Arg Asn Tyr Ser Val Asn Ile
 20 25 30

Arg Glu Glu Leu Lys Leu Ala Asn Val Val Phe Phe Pro Arg Cys Leu
 35 40 45

Leu Val Gln Arg Cys Gly Gly Asn Cys Gly Cys Gly Thr Val Lys Leu
 50 55 60

Glu Ser Cys Thr Cys Asn Ser Gly Lys Thr Val Lys Lys Tyr His Glu
 65 70 75 80

Val Leu Gln Phe Glu Pro Gly His Ile Lys Arg Arg Gly Arg Ala Lys
 85 90 95

Thr Met Ala Leu Val Asp Ile Gln Leu Asp His His Glu Arg Cys Asp
 100 105 110

Cys Ile Cys Ser Ser Arg Pro Pro Arg
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<210> 19

<211> 119

<212> PRT

<213> Homo sapiens

<400> 19

Arg Asp Glu Thr Ile Gln Val Lys Gly Asn Gly Tyr Val Gln Ser Pro
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Arg Phe Pro Asn Ser Tyr Pro Arg Asn Leu Leu Leu Thr Trp Arg Leu
 20 25 30

His Ser Gln Glu Asn Thr Arg Ile Gln Leu Val Phe Asp Asn Gln Phe
 35 40 45

Gly Leu Glu Glu Ala Glu Asn Asp Ile Cys Arg Tyr Asp Phe Val Glu
 50 55 60

Val Glu Asp Ile Ser Glu Thr Ser Thr Ile Ile Arg Gly Arg Trp Cys
 65 70 75 80

Gly His Lys Glu Val Pro Pro Arg Ile Lys Ser Arg Thr Asn Gln Ile
 85 90 95

Lys Ile Thr Phe Lys Ser Asp Asp Tyr Phe Val Ala Lys Pro Gly Phe
 100 105 110

Lys Ile Tyr Tyr Ser Leu Leu
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<210> 20

<211> 113

<212> PRT

<213> Homo sapiens

<400> 20

Cys Gly Glu Thr Leu Gln Asp Ser Thr Gly Asn Phe Ser Ser Pro Glu
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Tyr Pro Asn Gly Tyr Ser Ala His Met His Cys Val Trp Arg Ile Ser
 20 25 30

Val Thr Pro Gly Glu Lys Ile Ile Leu Asn Phe Thr Ser Leu Asp Leu
 35 40 45

Tyr Arg Ser Arg Leu Cys Trp Tyr Asp Tyr Val Glu Val Arg Asp Gly
 50 55 60

Phe Trp Arg Lys Ala Pro Leu Arg Gly Arg Phe Cys Gly Ser Lys Leu
 65 70 75 80

Pro Glu Pro Ile Val Ser Thr Asp Ser Arg Leu Trp Val Glu Phe Arg
 85 90 95

Ser Ser Ser Asn Trp Val Gly Lys Gly Phe Phe Ala Val Tyr Glu Ala
 100 105 110

Ile

<210> 21
 <211> 112
 <212> PRT
 <213> Homo sapiens

<400> 21
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 Tyr Pro Asp Asp Tyr Arg Pro Ser Lys Val Cys Ile Trp Arg Ile Gln
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 Val Ser Glu Gly Phe His Val Gly Leu Thr Phe Gln Ser Phe Glu Ile
 35 40 45
 Glu Arg Met Asp Ser Cys Ala Tyr Asp Tyr Leu Glu Val Arg Asp Gly
 50 55 60
 His Ser Glu Ser Ser Thr Leu Ile Gly Arg Tyr Cys Gly Tyr Glu Lys
 65 70 75 80
 Pro Asp Asp Ile Lys Ser Thr Ser Ser Arg Leu Trp Leu Lys Phe Val
 85 90 95
 Ser Asp Gly Ser Ile Asn Lys Ala Gly Phe Ala Val Asn Phe Phe Lys
 100 105 110

<210> 22
 <211> 113
 <212> PRT
 <213> Homo sapiens

<400> 22
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 20 25 30
 Ala Pro Thr Gln Tyr Arg Ile Ser Leu Gln Phe Asp Phe Phe Glu Thr
 35 40 45

Glu Gly Asn Asp Val Cys Lys Tyr Asp Phe Val Glu Val Arg Ser Gly
 50 55 60
 Leu Thr Ala Asp Ser Lys Leu His Gly Lys Phe Cys Gly Ser Glu Lys
 65 70 75 80
 Pro Glu Val Ile Thr Ser Gln Tyr Asn Asn Met Arg Val Glu Pro Lys
 85 90 95
 Ser Asp Asn Thr Val Ser Lys Lys Gly Phe Lys Ala His Phe Phe Ser
 100 105 110

Glu

<210> 23
 <211> 113
 <212> PRT
 <213> Homo sapiens

<400> 23
 Gly Asp Thr Ile Lys Ile Glu Ser Pro Gly Tyr Leu Thr Ser Pro Gly
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 Tyr Pro His Ser Tyr His Pro Ser Glu Lys Cys Glu Trp Leu Ile Gln
 20 25 30
 Ala Pro Asp Pro Tyr Gln Arg Ile Met Ile Asn Phe Asn Pro His Phe
 35 40 45
 Asp Leu Glu Asp Arg Asp Cys Lys Tyr Asp Tyr Val Glu Val Phe Asp
 50 55 60
 Gly Glu Asn Glu Asn Gly His Phe Arg Gly Lys Phe Cys Gly Lys Ile
 65 70 75 80
 Ala Pro Pro Pro Val Val Ser Ser Gly Pro Phe Leu Phe Ile Lys Phe
 85 90 95
 Val Ser Asp Tyr Glu Thr His Gly Ala Gly Phe Ser Ile Arg Tyr Glu
 100 105 110
 Ile

<210> 24

<211> 119
 <212> PRT
 <213> Homo sapiens

<400> 24
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 20 25 30
 Ala Pro Lys Met Ser Glu Ile Ile Leu Glu Phe Glu Ser Phe Asp Leu
 35 40 45
 Glu Pro Asp Ser Asn Pro Pro Gly Gly Met Phe Cys Arg Tyr Asp Arg
 50 55 60
 Leu Glu Ile Trp Asp Gly Phe Pro Asp Val Gly Pro His Ile Gly Arg
 65 70 75 80
 Tyr Cys Gly Gln Lys Thr Pro Gly Arg Ile Arg Ser Ser Ser Gly Ile
 85 90 95
 Leu Ser Met Val Phe Tyr Thr Asp Ser Ala Ile Ala Lys Glu Gly Phe
 100 105 110
 Ser Ala Asn Tyr Ser Val Leu
 115

<210> 25
 <211> 15
 <212> PRT
 <213> Homo sapiens

<400> 25
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 1 5 10 15

<210> 26
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 <212> DNA
 <213> Homo sapiens

<400> 26
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<210> 27
<211> 20
<212> DNA
<213> Homo sapiens

<400> 27
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<210> 28
<211> 27
<212> DNA
<213> Homo sapiens

<400> 28
ccatcctaatac gactcact atagggc

27

<210> 29
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<212> DNA
<213> Homo sapiens

<400> 29
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<210> 30
<211> 26
<212> DNA
<213> Homo sapiens

<400> 30
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26

<210> 31
<211> 22
<212> DNA
<213> Homo sapiens

<400> 31
tgctctagat cgagggtggtc tt

22

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/26462

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C07K 14/49, 14/475; C12N 15/00

US CL : 435/69.1; 536/23.5; 530/350

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. :

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Genbank, EST, Swissprot, Medline, USPatfull
search terms: PDGF-D, VEGF, CUB

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 5,607,918 A (ERIKSSON et al) 04 March 1997(04.03.97), col.13-14, SEQ ID No:2 at position 39-51.	10, 30
A	WO 96/27007 A1 (AMRAD OPERATIONS PTY. LTD.) 06 September 1996(06.09.96), see page 41, SEQ ID No:4 at position 70-82.	10, 30
A	Database GENCORE on EST, AN AA488780, NCI-CGAP, 'National Cancer Institute, Cancer Genome Anatomy Project (CGAP), Tumor Gene Index.' Gene Sequence, 15 August 1997.	1-40, 50, 52-60, 66,67
A	Database GENCORE on EST, AN AA488996, NCI-CGAP, 'National Cancer Institute, Cancer Genome Anatomy Project (CGAP), tumor Gene Index.' Gene Sequence, 15 August 1997.	1-40, 50, 52-60, 66, 67



Further documents are listed in the continuation of Box C.



See patent family annex.

Special categories of cited documents:	
A document defining the general state of the art which is not considered to be of particular relevance	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
E earlier document published on or after the international filing date	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	*Z* document member of the same patent family

Date of the actual completion of the international search

23 MARCH 2000

Date of mailing of the international search report

25 APR 2000

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/26462

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	Database GENCORE on EST, AN AA736766, NCI-CGAP, 'National Cancer Institute, Cancer Genome Anatomy Project (CGAP), Tumor Gene Index.' Gene Sequence, 23 January 1998.	1-40, 50, 52-60, 66, 67
A	Database GENCORE on EST, AN AQ041639, ADAMS et al. 'Use of a random BAC End Sequence Database for Sequence-Ready Map Building (1998).' Gene Sequence, 14 July 1998.	1-40, 50, 52-60, 66, 67

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/26462

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:
Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☒ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
1-40, 50, 52-60, 66, 67
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest



The additional search fees were accompanied by the applicant's protest.



No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/26462

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

- Group I, claims 1-40, 50, 55-60, and 66, drawn to nucleic acid molecules having SEQ ID No:3, 5 and 7, vectors, host cells, proteins having SEQ ID No:4, 6 and 8, and method of making a polypeptide.
- Group II, claims 41-42, drawn to a method of amplifying a polynucleotide.
- Group III, claims 43-49, drawn to antibodies.
- Group IV, claim 51, drawn to a method of stimulating growth.
- Group V, claims 52-54 and 67, drawn to a method of producing an activated truncated form of PDGF-D.
- Group VI, claim 61, drawn to a method of inducing PDGF beta receptor activation.
- Group VII, claim 62, drawn to method of inhibiting tumor growth.
- Group VIII, claim 63, drawn to a method of identifying types of tumors.
- Group IX, claims 64-65, drawn to method of identifying a PDGF-D antagonist.
- Group X, claim 68, drawn to a method of inhibiting tissue remodeling.

The inventions listed as Groups I-X do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Pursuant to 37 C.F.R. d), the ISA/US considers that where multiple products and processes are claimed, the main invention shall consist of the first invention of the category first mentioned in the claims and the first recited invention of each of the other categories related thereto. Accordingly, the main invention (Group I) comprises the first recited product, a polynucleotide of SEQ ID No: 3,5,7 encoding hPDGF-D, a vector, a host cell, a method of producing a hPDGF-D protein of SEQ ID No:4, 6 and 8 and hPDGF-D protein of SEQ ID No:4,6 and 8. Note that there is no method of making the polynucleotide. Further, pursuant to 37 C.F.R. 1.45(d), the ISA/US considers that any feature which the subsequently products of Group III and additional methods of Groups II and IV-X share with the main invention does not constitute a special technical feature within the meaning of PCT Rule 13.2 and that each of such products and methods accordingly defines a separate invention.